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Ecological Modelling

## Multi-site Calibration and Validation of a Net Ecosystem Carbon Exchange Model for Croplands

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## 1 ABSTRACT

2 Croplands play an important role in the carbon budget of many regions. However, the  
3 estimation of their carbon balance remains difficult due to diversity and complexity of the  
4 processes involved. We report the coupling of a one-dimensional soil water, heat, and CO<sub>2</sub>  
5 flux model (SOILCO<sub>2</sub>), a pool concept of soil carbon turnover (RothC), and a crop growth  
6 module (SUCROS) to predict the net ecosystem exchange (NEE) of carbon. The coupled  
7 model, further referred to as AgroC, was extended with routines for managed grassland as  
8 well as for root exudation and root decay. In a first step, the coupled model was applied to  
9 two winter wheat sites and one upland grassland site in Germany. The model was calibrated  
10 based on soil water content, soil temperature, biometric, and soil respiration measurements for  
11 each site, and validated in terms of hourly NEE measured with the eddy covariance technique.  
12 The overall model performance of AgroC was sufficient with a model efficiency above 0.78  
13 and a correlation coefficient above 0.91 for NEE. In a second step, AgroC was optimized with  
14 eddy covariance NEE measurements to examine the effect of different objective functions,  
15 constraints, and data-transformations on estimated NEE. It was found that NEE showed a  
16 distinct sensitivity to the choice of objective function and the inclusion of soil respiration data  
17 in the optimization process. In particular, both positive and negative day- and nighttime fluxes  
18 were found to be sensitive to the selected optimization strategy. Additional consideration of  
19 soil respiration measurements improved the simulation of small positive fluxes remarkably.  
20 Even though the model performance of the selected optimization strategies did not diverge  
21 substantially, the resulting cumulative NEE over simulation time period differed substantially.  
22 Therefore, it is concluded that data-transformations, definitions of objective functions, and  
23 data sources have to be considered cautiously when a terrestrial ecosystem model is used to  
24 determine NEE by means of eddy covariance measurements.

1    Keywords: AgroC, soil respiration, carbon balance, winter wheat, grassland, NEE

## 1. Introduction

Terrestrial ecosystems play an important role in the global carbon cycle. Photosynthesis by vegetation and respiration from autotrophic and heterotrophic organisms represent the two major carbon fluxes between atmosphere and terrestrial biosphere. Terrestrial ecosystems store large amounts of carbon, and especially soils contain about twice as much carbon as the atmosphere (Rustad et al., 2000). Over 37% of the world's landmass is agricultural land (FAO Statistical Yearbook, 2014). Thus, carbon fluxes in agroecosystems constitute a significant part of the global carbon cycle. The quantification and prediction of terrestrial carbon sinks and sources and their dynamics, variabilities, and controls are of major importance with regards to climate change research and to optimization of management strategies affecting the ecosystem's carbon budget (e.g., Baldocchi, 2003; Kuzyakov, 2006; Subke et al., 2006). The net ecosystem exchange (NEE) of carbon dioxide and its two components, gross primary production (GPP) and terrestrial ecosystem respiration (TER), are of particular interest (Suleau et al., 2011; Sus et al., 2010). The total CO<sub>2</sub> efflux from soils, one of the major compartments of TER (Moureaux et al., 2008; Suleau et al., 2011), derives from decomposition of soil organic matter and dead plant material by microorganisms, from direct root respiration, and from microbial respiration of root exudates and rhizodepositions (Kuzyakov, 2006; Kuzyakov and Domanski, 2000). In this study, we consider the last two CO<sub>2</sub> sources as one sum, and refer to it as "rhizosphere respiration".

NEE is increasingly being monitored using the eddy covariance (EC) technique, which provides information on net carbon fluxes for a relatively large area with a high temporal resolution (Baldocchi, 2003). This allows to investigate the relation between CO<sub>2</sub> efflux and weather conditions or crop development stages (Sus et al., 2010). Due to methodological and technical constraints, significant gaps occur in high-quality EC data, which prohibits direct computation of annual NEE. Gap-filling methods (e.g., Reichstein et al., 2005) and their

application with meteorological and EC data overcome this limitation, but e.g., they cannot be used for predictive modeling of carbon balances addressing climate change effects. Alternatively, terrestrial ecosystem models with a physical description of processes in the agroecosystem can be used to assess annual NEE sums. An additional advantage of such models is that they allow to quantify interrelations and feedbacks in biogeochemical processes and fluxes of agricultural systems. Mechanistic models like ORCHIDEE-STICS (de Noblet-Ducoudré et al., 2004), DNDC (Li et al., 2005), or SPAC (Sus et al., 2010) were developed for this purpose and have been successfully applied in a number of studies (e.g., Sus et al., 2010; Wattenbach et al., 2010; Wu et al., 2009; Yuan et al., 2012). In most of these studies, the carbon assimilation by plants was captured well by the models, but a significant bias in the simulation of the respiratory fluxes was observed. This inevitably causes systematic errors in the estimation of the overall carbon balance. An improved representation of processes linked to respiration may help to decrease systematic errors and in combination with soil respiration ( $R_{\text{soil}}$ ) measurements, it may help to reduce the uncertainty in the estimation of annual NEE. For this purpose, we coupled a one-dimensional soil water, heat, and CO<sub>2</sub> flux model (SOILCO<sub>2</sub>; Šimůnek and Suarez, 1993), a pool concept of soil carbon turnover (RothC; Coleman and Jenkinson, 2008), and a crop growth module (SUCROS; Spitters et al., 1989). In addition, the coupled model, further referred to as AgroC, was extended with routines for root exudation, root decay, as well as for a managed grassland system. The main motivation for the coupling was a more detailed representation of sources and locations of CO<sub>2</sub> production, the gas transport in the soil, and the fluxes in the ecosystem. Various sources of measured data are available for validation, calibration, evaluation, and structural improvement of terrestrial ecosystem models. In the last decade, substantial progress has been made in implementing model-data fusion techniques to make optimal use of available measurements (e.g., Richardson et al., 2010; Sus et al., 2010; Trudinger et al.,

2007; Wu et al., 2009; Yuan et al., 2012). Such model-data fusion techniques, including calibration techniques, require the formulation and minimization of an objective function that quantifies the mismatch between model predictions and observations (Evans, 2003; Herbst et al., 2008; Wang et al., 2009). Detailed measurements of biotic and abiotic processes and fluxes allow to improve process models on various spatiotemporal scales, and to verify model assumptions, parameters, and performance (Richardson et al., 2010; Williams et al., 2009; Yuan et al., 2012). However, the use of multiple objective functions or constraints in model calibration may be challenging because of the need to combine measurements with variable spatial scale, temporal scale, magnitude, and uncertainty. For example, optimizing the simulation regarding one data source (e.g., NEE) can lead to a low model performance (trade-off) regarding another data source (e.g., heterotrophic soil respiration) (Richardson et al., 2010). Other important decisions to be made before model calibration include the selection and appropriate weighting of observations, the choice of an optimization algorithm (Trudinger et al., 2007), and the selection of model parameters being altered during calibration (Wu et al., 2009). These decisions differ between model studies, which will influence the results of NEE predictions (Evans, 2003; Trudinger et al., 2007).

The main goal of this study is to present the mechanistic model AgroC and to evaluate its model performance simulating biophysical processes and interactions in agroecosystems. In a first step, AgroC was calibrated with soil moisture, soil temperature, biometric, and soil CO<sub>2</sub> flux measurements of three test sites in Germany cropped with winter wheat, barley, or grass. After calibration, it was evaluated how well AgroC simulates the hourly NEE through comparison with EC measurements. In the next step, we optimized the AgroC model using EC measurements by estimating plant and  $R_{soil}$  parameters. In addition, we evaluated how joint use of EC and  $R_{soil}$  measurements in the calibration affected the estimated cumulative

NEE and model performance. Finally, we evaluated the effect of data-transformation (e.g., log-transformation) on the model results with a focus on estimated NEE.

## **2. Materials and Methods**

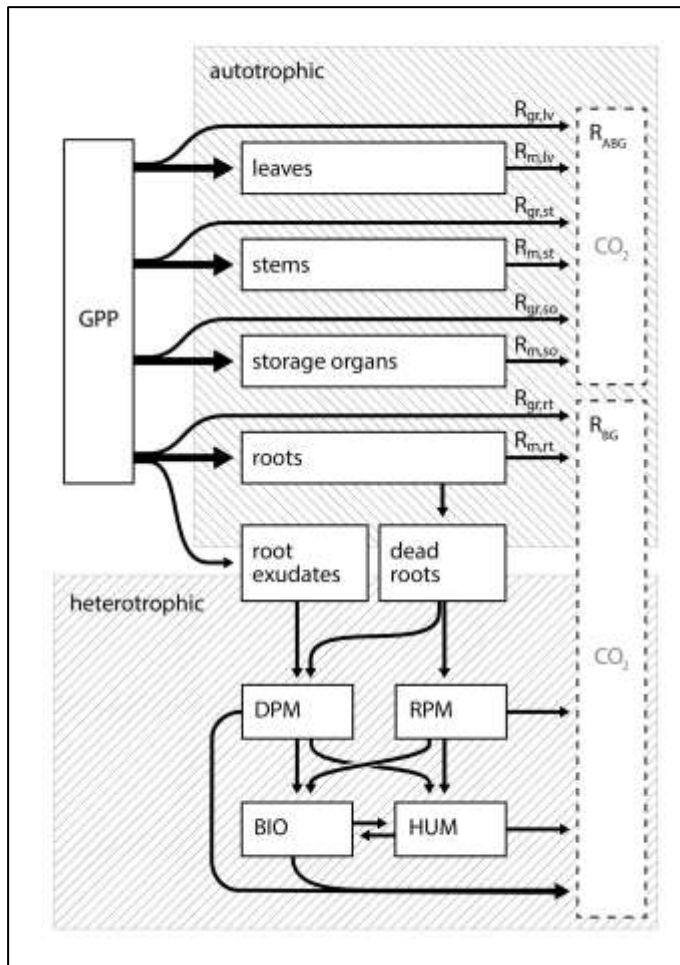
### *2.1. The AgroC Model*

AgroC is a coupled model developed from the SOILCO<sub>2</sub>/RothC model (Herbst et al., 2008) and the SUCROS model for crop growth (Spitters et al., 1989). The SOILCO<sub>2</sub>/RothC model simulates vertical water, heat, and CO<sub>2</sub> fluxes in a soil column, and the source term of heterotrophic respiration over soil depth and time, which is given by the turnover of depth-specific carbon pools (Coleman and Jenkinson, 2008; Šimůnek and Suarez, 1993; Šimůnek et al., 1996). The carbon turnover rates depend on the soil water content and temperature. The SOILCO<sub>2</sub>/RothC model was validated in several laboratory and field studies (Bauer et al., 2008, 2012; Herbst et al., 2008; Palosuo et al., 2012; Weihermüller et al., 2009). The extension with SUCROS is expected to allow for an improved simulation of the soil autotrophic respiration source term, since temporal development of root growth and related growth and maintenance respiration is simulated by SUCROS in a mechanistic way. In addition, AgroC was extended with routines for root exudation and root decay. Furthermore, this coupled model allows closing the one-dimensional carbon balance and to estimate NEE, since carbon assimilation as well as organ-specific growth and maintenance respiration are now included. Figure 1 provides a summary of the carbon cycling in AgroC. Moreover, routines for the simulation of managed grassland were implemented in AgroC following the sink/source approach suggested by Schapendonk et al. (1998) for the grassland productivity model LINGRA.

AgroC was adapted to work with an hourly time step. The coupled SOILCO<sub>2</sub>/RothC model allows the use of user-specified length and time units, whereas the SUCROS module uses



1 fixed units. For the coupled AgroC model, we preserved the flexibility in terms of length ([L])  
2 and time units ([T]), but we kept the fixed mass and area units (kg, ha) of the original  
3 SUCROS code. Further information about the coupling and the modifications to the original  
4 models regarding the hourly time step, the water fluxes, the carbon fluxes, and the grassland  
5 routines are given in the Appendix A.



*Fig. 1:*

Carbon fluxes and partitioning in AgroC. Gross primary production (GPP) is partitioned to the different plant organs, leaves (subscript lv), stems (st), storage organs (so), and roots (rt).  $CO_2$  is lost due to growth ( $R_{gr}$ ) and maintenance respiration ( $R_m$ ). The sum of these autotrophic  $CO_2$  source terms by the shoot organs account for the above-ground respiration ( $R_{ABG}$ ). Carbon and  $CO_2$  is added to the soil profile by autotrophic root respiration, root exudates, and dead roots. The latter two are transferred to the decomposable and resistant plant material pool (DPM, RPM) of the RothC model and decomposed. The heterotrophic  $CO_2$  source term consists of microbial decomposition of those and further soil organic matter pools (humified organic matter HUM, microbial biomass BIO). The root respiration and the heterotrophic components are part of the below-ground respiration ( $R_{BG}$ ).

## 2.2. Study Sites and Data Availability

AgroC was applied to three experimental sites in the western part of Germany: Selhausen and Merzenhausen, both located in the southern part of the Lower Rhine Embayment (Schmidt et al., 2012; Stadler et al., 2015), and Rollesbroich, located in the low mountain range Eifel (Gebler et al., 2015). The dominant land use at the first two test sites is cropland. Rollesbroich is a managed grassland site, which is mown three times per year (Borchard et al., 2015). All three study sites are included in the Terrestrial Environmental Observatories (TERENO) network of highly instrumented field sites (Zacharias et al., 2011). An overview of soil properties, meteorological conditions, and crop management is given in Tables 1 and A.1 for all three sites.

At the two cropland sites, EC and ancillary environmental measurements were conducted in the center of the agricultural fields. Measurements of NEE, latent heat, wind components, global radiation, air temperature, soil (surface) temperature at a depth of -1 cm, precipitation, and relative humidity were collected. A detailed description of the sites, measurement setup, EC post-processing, and footprint modelling is given by Schmidt et al. (2012), Graf et al. (2013), Post et al. (2015), Mauder et al. (2013) and Kormann and Meixner (2001). Soil water content and soil temperature were measured in various depths at several soil profiles per site. Biometric measurements were carried out bi-weekly to monitor crop development, and  $R_{\text{soil}}$  data were obtained with closed-chamber measurements during summer (Prolingheuer et al., 2014; Schmidt et al., 2012; Stadler et al., 2015). Prolingheuer et al. (2014) also measured the heterotrophic contribution to the  $\text{CO}_2$  flux by root exclusion experiments at 61 sample points at the Selhausen test site.

In Rollesbroich, the EC tower was placed between two neighboring grasslands (A and B) with different management in terms of mowing dates. Thus, measured fluxes were dominated by one of the two grasslands depending on the wind direction and the resulting flux footprint

1 distribution. Data processing was similar to the two agricultural fields. Borchard et al. (2015)  
2 conducted detailed surveys of the Rollesbroich site. At 21 sample points in grassland A, soil  
3 samples were taken, and total LAI and harvested dry matter were also determined during the  
4 growing season. Eleven of the sampling points were mown following the management of  
5 grassland A, and the remaining 10 points were sampled following the management of  
6 grassland B.  $R_{\text{soil}}$  was again determined from closed-chamber measurements during summer.  
7 Soil moisture, soil temperature, and  $\text{CO}_2$  concentration in several depths were observed at  
8 three profiles near the EC tower.

*Tab. 1:*

Site-specific characteristics, meteorological conditions, and crop management (WW: Winter wheat; WB: winter barley; GL: grassland) (Borchard et al., 2015; Gebler et al., 2015; Prolingheuer et al., 2014; Schmidt et al., 2012; Séquaris et al., 2013; Stadler et al., 2015).

	Selhausen	Merzenhausen	Rollesbroich
<b>Site characteristics</b>			
coordinates	50°52'14''N, 6°26'59''E	50°55'47''N, 6°17'49''E	50°37'19''N, 6°18'15''E
elevation (m a.s.l.)	103	93	515
soil type*	Luvisol	Luvisol	Cambisol
soil texture	silt loam	silt loam	silty clay
<b>Climate conditions</b>			
mean annual temperature (°C)	9.9	9.9	7.7
annual precipitation (mm)	698	698	1033
<b>Simulation period</b>	Oct 2008 - Dec 2009	Oct 2011 - Dec 2014	Jan 2013 - Dec 2013
<b>Land management</b>			
crop sequence	WW tilled every autumn	WW - WW - WB tilled every autumn	GL mowed 3x annually

\*according to soil taxonomy of the FAO (I.U.S.S. Working Group WRB, 2006)

### 2.3. Model Setup and Initialization

AgroC requires gap-filled meteorological data (air temperature, soil surface temperature, precipitation, solar radiation, and potential grass reference evapotranspiration), plant-specific parameters, and soil characteristics. Potential grass reference evapotranspiration was estimated with the Penman-Monteith approach according to the FAO guidelines (Allen et al., 1998). Plant-specific parameters for cereals and grass were mainly taken from literature (e.g., Boons-Prins et al., 1993; Gonzales et al., 1989; Goudriaan et al., 1997; Kuzyakov and Domanski, 2000; Parsons, 1988; Parsons and Robson, 1981; Prud'homme et al., 1992; Schapendonk et al., 1998; Spitters et al., 1989; Swinnen et al., 1995; Vanclooster et al., 1995; van Keulen et al., 1997). These plant parameters have been extensively used in other simulation studies with the models SUCROS and LINGRA. Root biomass measurements were not available, thus the proportion of the root system (root/shoot ratio) was also derived from literature (e.g., Bolinder et al., 1997, 2002; López et al., 2013).

In AgroC appropriate boundary conditions have to be specified for CO<sub>2</sub>, water, and heat flow at the top and bottom of the simulation domain. The upper boundary condition for CO<sub>2</sub> flow was the atmospheric concentration of 0.038%. Meteorological measurements were used to describe the upper boundary for water and heat flux. Soil profile characteristics were available from Séquaris et al. (2013), Herbst et al. (2005), and Borchard et al. (2015) for Selhausen, Merzenhausen, and Rollesbroich, respectively (Tab. A.1). The simulated profile depths varied from 1.0 to 1.2 m. A no-flow boundary was used at the bottom of the soil profile for heat and CO<sub>2</sub>. For water, a prescribed pressure head following a sine wave over the course of the year with a minimum in autumn was used as a Dirichlet boundary condition at the bottom of the simulation domain (Bauer et al., 2008; Scharnagl et al., 2011).

Initial carbon pool sizes were derived from measured soil organic carbon contents for each soil horizon. In Selhausen and Rollesbroich, measured soil carbon fractions were available

from previous studies (Bauer et al., 2012; Séquaris et al., 2013; Nils Borchard and Henning Schiedung, personal communication). For these two sites, initial pool sizes were calculated following Falloon et al. (1998), Skjemstad et al. (2004), and Zimmermann et al. (2007). For Merzenhausen, initial pool sizes were determined with pedotransfer functions according to Weihermüller et al. (2013), assuming a state of equilibrium. The reference temperature required for the estimation of the soil heterotrophic CO<sub>2</sub> source term was set to the mean annual temperature at each site.

#### 2.4. Model Calibration

In a first step, AgroC was calibrated with the downhill Nelder-Mead Simplex algorithm (Nelder and Mead, 1965), since only a small number of parameters were considered. The root mean square error (*RMSE*) between measurements and simulations was minimized. In addition, the Pearson product-moment correlation coefficient (*r*) and the model efficiency (*ME*) (Nash and Sutcliffe, 1970) were calculated as model quality criteria. A *ME* close to 1 indicates that the simulation describes the observations well without systematic bias. If *ME* is lower than 0, the mean of the observations is a better predictor than the simulations.

First, the soil hydraulic parameters were calibrated. Then, plant development and growth were adjusted. Here, mainly the plant development rate depending on temperature, the effectiveness of CO<sub>2</sub> assimilation, the partitioning factors of assimilates between the different plant organs, especially between shoot and root system, and the specific leaf area (conversion factor between plant dry matter and LAI) were modified (Tab. A.2).

CO<sub>2</sub> production in the soil profile was estimated in dependence of several physical processes and conditions. For soil temperature, we used the default reduction function of the SOILCO<sub>2</sub> model, which is a modified form of the Arrhenius relationship (Šimůnek and Suarez, 1993; Šimůnek et al., 1996). To describe the soil moisture dependency of respiration, we applied a

bell-shaped curve as suggested by Bauer et al. (2012), Moyano et al. (2012), and Skopp et al. (1990). The simulation of  $R_{\text{soil}}$  was improved by calibrating the reference temperature used in the temperature scaling function, the turnover rate of the RPM pool, and the parameters of the water reduction function. For Rollesbroich, soil  $\text{CO}_2$  concentration measurements in different depths were available, so the gaseous diffusion through the soil matrix could also be adjusted. Here, we implemented the gas diffusivity and transport model of Kristensen et al. (2010), which accounts for preferential diffusion through fractures and macropores in the soil matrix. Appendant parameters, the fracture porosity, the fracture porosity factor, and the matrix tortuosity factor, were adjusted.

After soil water, soil heat, and  $\text{CO}_2$  flux, as well as plant development were calibrated, we compared the NEE estimates with the EC measurements at each test site. NEE measurements were handled according to the quality assessment strategy suggested by Mauder et al. (2013), and only data with high quality was used for validation purposes (28% of data in Selhausen; 55% of data in Merzenhausen; 33% of data in Rollesbroich).

In a second step, several model runs were conducted where simulated NEE was optimized with EC measurements by estimating plant parameters (regarding the light use efficiency, the potential  $\text{CO}_2$  assimilation rate, their dependence on crop DVS and air temperature, and the biomass partitioning factors between shoot and root), and model parameters affecting  $R_{\text{soil}}$  (as above: reference temperature, turnover rate of RPM, and parameters of the water reduction function). Here, parameter calibration was conducted with the Shuffled Complex Evolution (SCE) algorithm (Duan et al., 1993), which is a global optimization strategy that was shown to be effective for a wide range of non-linear optimization problems. Two different objective functions were considered: (i) the *RMSE* and (ii) the sum of the *RMSE* and the *Bias*. The former was calculated on the basis of various data expressions (instantaneous data, cumulative data, or instantaneous log-transformed data). Additional calibrations were



1 conducted that not only considered NEE data for the optimization, but also measurements of  
2  $R_{\text{soil}}$ . Therefore, we considered a total of eight different calibration strategies (see Tab. 2).  
3 Because of the different magnitude of NEE and  $R_{\text{soil}}$  (and resulting misfits), the error was  
4 transformed by division with the respective observed mean flux (with the exception of  $NEE_{BSc}$   
5 approach). For each test site, these eight calibrations were conducted to examine the  
6 sensitivity of estimated cumulative NEE to the different objective functions and to the  
7 inclusion of  $R_{\text{soil}}$  measurements. Estimated cumulative NEE based on each optimization  
8 strategy was compared to the well-established gap-filling method by Reichstein et al. (2005),  
9 which is based on linear regressions between EC measurements and physical drivers.

1 *Tab. 2:*  
2 Applied optimization strategies and their objective functions, used data streams and data  
3 transformation (*obs\_N*: NEE observation; *sim\_N*: NEE simulation; *obs\_R*: Rsoil observation;  
4 *sim\_R*: Rsoil simulation).  
5

label	objective function	data streams	data transformation	obs or sim
$NEE_{inst}$			instantaneous	with $x_i$
$NEE_{Cum}$	$E = \sqrt{\frac{1}{n} \sum_{i=1}^n (obs\_N_i - sim\_N_i)^2}$	NEE	cumulative	$x_i = \sum_{j=1}^i x_j$
$NEE_{Log}$			log-transformed	$x_i = \ln(x_i +  min  + 1)$
<b>RMSE</b>				
$NEE_{inst} + R_{soil}$			instantaneous	$x_i$
$NEE_{Cum} + R_{soil}$	$E = \frac{\sqrt{\frac{1}{n} \sum_{i=1}^n (obs\_N_i - sim\_N_i)^2}}{\frac{1}{n} \sum_{i=1}^n obs\_N_i} + \frac{\sqrt{\frac{1}{m} \sum_{j=1}^m (obs\_R_j - sim\_R_j)^2}}{\frac{1}{m} \sum_{j=1}^m obs\_R_j}$	NEE and $R_{soil}$	cumulative	$x_i = \sum_{j=1}^i x_j$ *
$NEE_{Log} + R_{soil}$			log-transformed	$x_i = \ln(x_i +  min  + 1)$
<b>RMSE + Bias</b>				
$NEE_{BSc}$	$E = \sqrt{\frac{1}{n} \sum_{i=1}^n (obs\_N_i - sim\_N_i)^2 + \left  \frac{1}{n} \sum_{i=1}^n (obs\_N_i - sim\_N_i) \right }$	NEE	instantaneous	$x_i$
$NEE_{BSc} + R_{soil}$	$E = \sqrt{\frac{1}{n} \sum_{i=1}^n (obs\_N_i - sim\_N_i)^2 + \left  \frac{1}{n} \sum_{i=1}^n (obs\_N_i - sim\_N_i) \right } + \sqrt{\frac{1}{m} \sum_{j=1}^m (obs\_R_j - sim\_R_j)^2}$	NEE and $R_{soil}$	instantaneous	$x_i$

\* only applied to NEE data,  $R_{soil}$  data was used instantaneous.

### 3. Results and Discussion

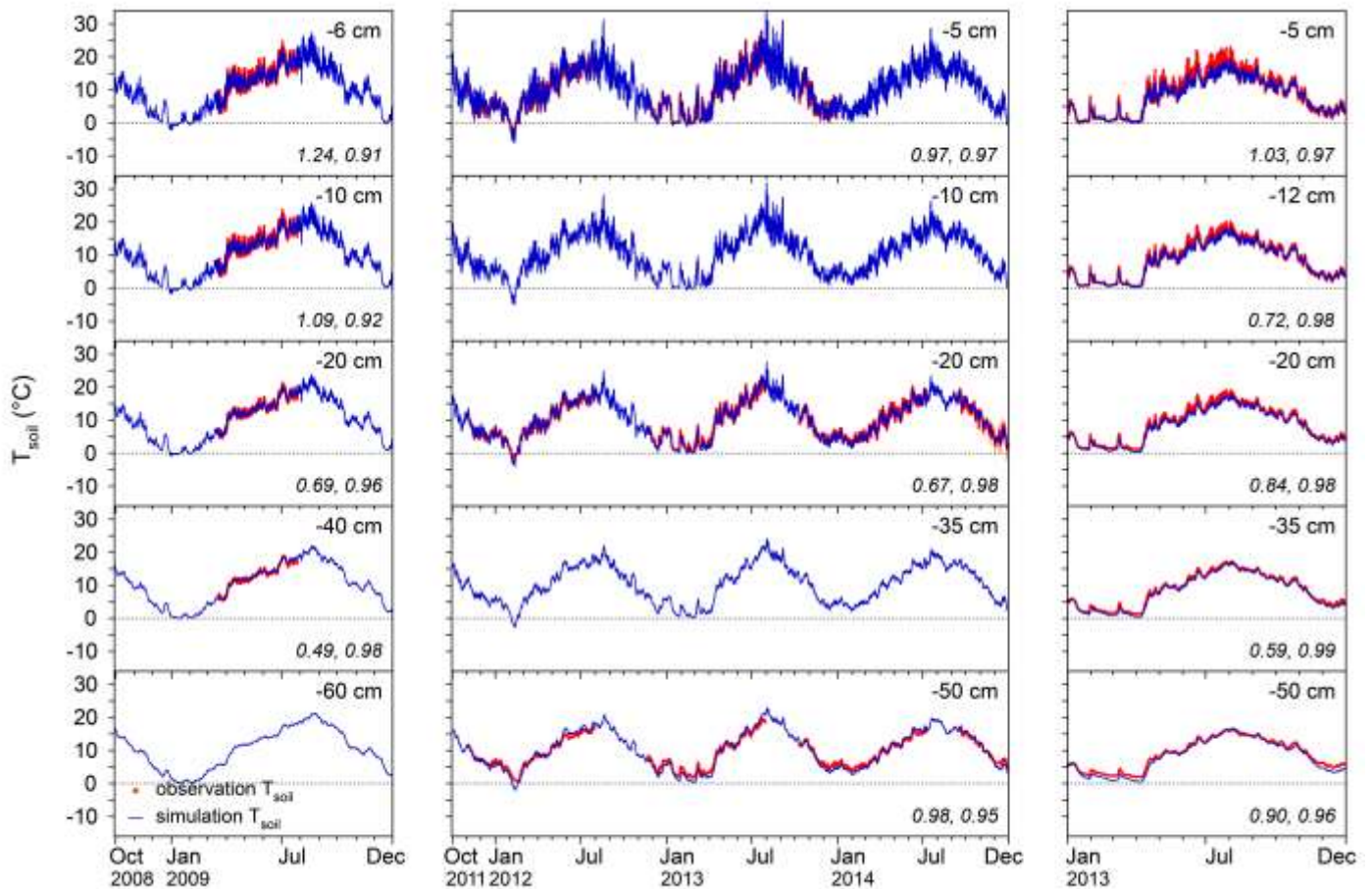
#### 3.1. Calibration and Validation of AgroC

##### *Soil Temperature and Water Content*

All simulations described measured soil temperature very well using the default settings. The *RMSE* was below  $1.0^{\circ}\text{C}$  and the *ME* larger than 0.93 when measurements for all depths and sites were considered (see Fig. 2).

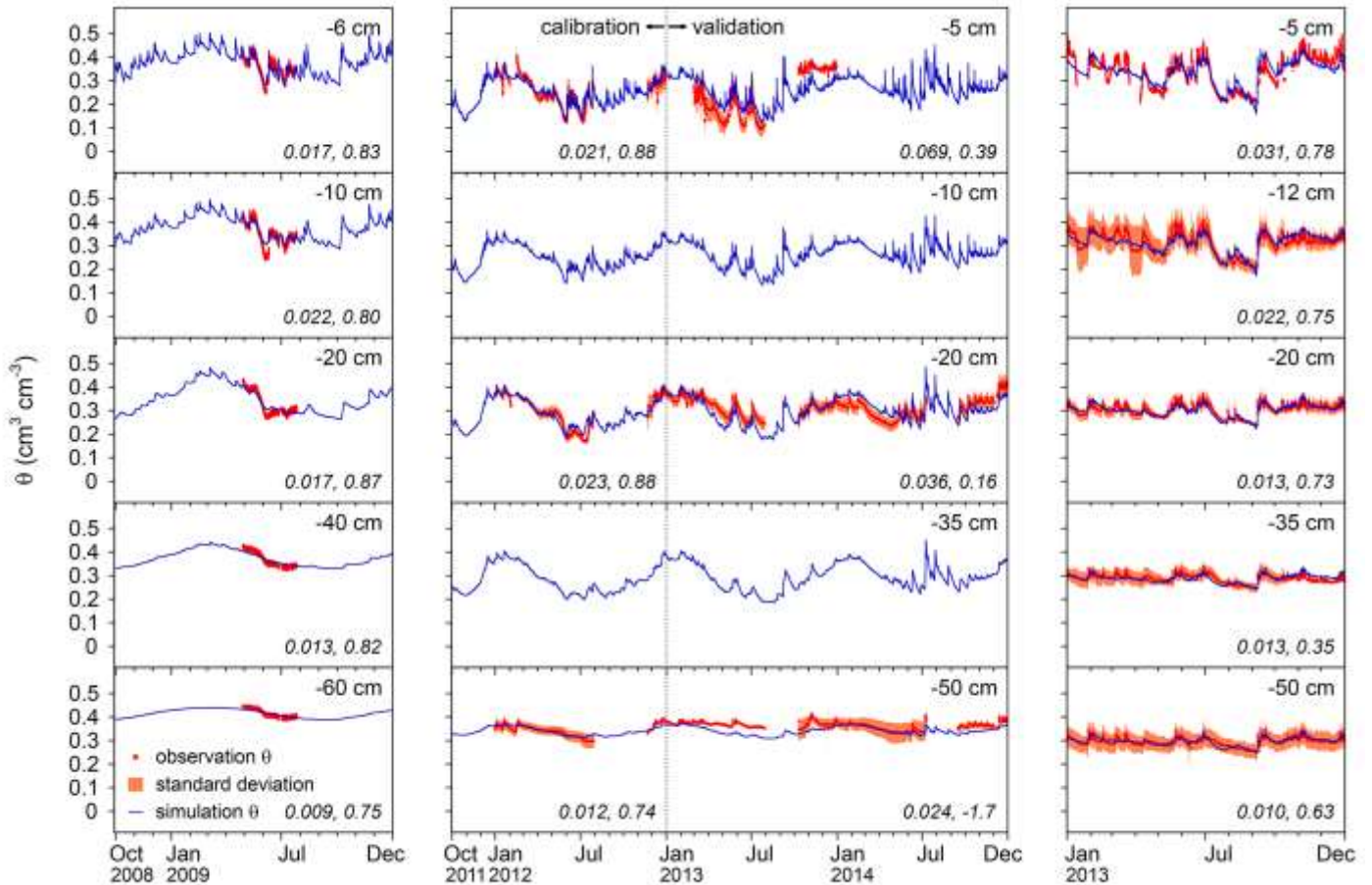
After calibration, the soil moisture dynamics were reproduced well by the AgroC model (Fig. 3). Estimated soil hydraulic parameters are summarized in Table A.1. The *RMSE* was below  $0.020 \text{ cm cm}^{-3}$ , the *ME* above 0.74 and the *r* above 0.86 for all sites and profile depths.

For Merzenhausen, the model was calibrated for 2012 and the following two years were used for validation. The performance of the model decreased for the validation period, but overall dynamics were still reproduced well (Fig. 3). Some near-surface peaks in soil moisture were not captured by the model, which is probably related to inaccuracies in the meteorological data used for the upper boundary condition. Furthermore, static hydraulic properties were assumed for the AgroC simulations, which is a simplification because the hydraulic properties of managed topsoils are typically variable due to ploughing, seedbed preparation, and subsequent re-compaction. For the Rollesbroich site, soil moisture simulations at -5 cm differed from the observations during winter. This is partly related to the presence of a snow cover, which results in delayed infiltration not represented in the model, and frozen soil, which affects soil water content measurements with the dielectric sensors used in this study.



*Fig. 2:*

Observed (dots; orange area: standard deviation) and simulated (lines) soil temperature ( $T_{\text{soil}}$ ) in several depths in Selhausen (left), Merzenhausen (middle), and Rollesbroich (right). Root mean square error (RMSE) and model efficiency (ME) (in this order) are given for each soil depth and location.



*Fig. 3:*

Observed (dots; orange area: standard deviation) and simulated (lines) soil water content ( $\theta$ ) at various depths in Selhausen (left), Merzenhausen (middle), and Rollesbroich (right). Root mean square error (RMSE) and model efficiency (ME) (in this order) are given for each soil depth and location. In Merzenhausen, RMSE and ME are given for the calibration (until end of 2012) and the validation period.

## 1 *Crop Development and Growth*

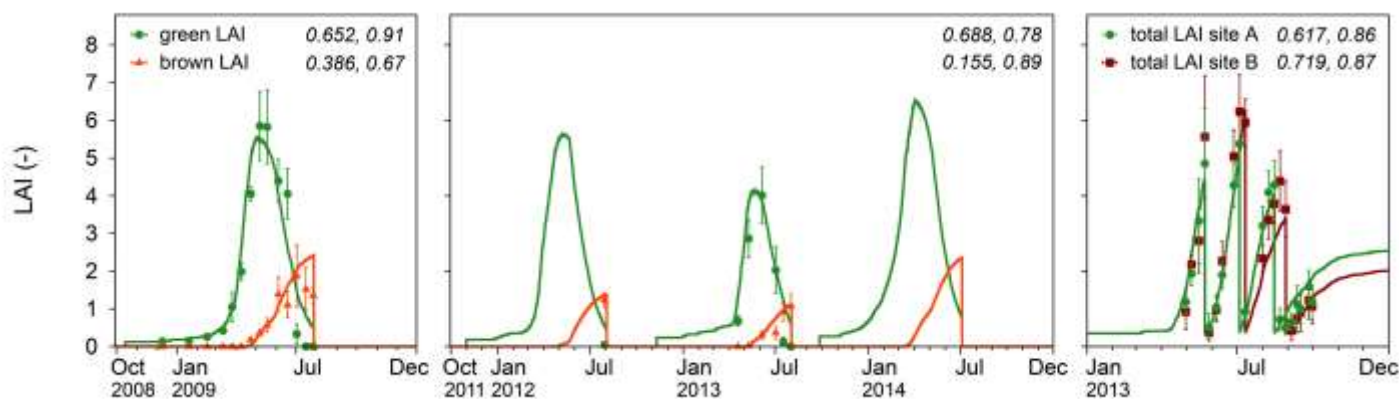
2 Without calibration, simulated crop development and dry matter accumulation over time were  
 3 already close to the observations (not shown). For further improvement, plant-specific  
 4 parameters were manually adjusted (Fig. 4, 5). In general, the assimilation rate, the fraction of  
 5 the root biomass, and the specific leaf area were increased for all crops at all test sites. In  
 6 Table A.2 in the appendix, the most relevant plant parameters are summarized. For total LAI,  
 7 the lowest *ME* was 0.63, *RMSE* was lower than 0.82 ha ha<sup>-1</sup>, and *r* was larger than 0.93 for all  
 8 sites. Site-specific errors for green and brown LAI are provided in Figure 4. As can be seen,  
 9 green LAI was well reproduced over the growing season, while the course of brown LAI was  
 10 simulated less well. As indicated by the *ME* in Figure 5, the simulation of dry matter was  
 11 adequate too, especially for winter wheat in Selhausen. However, the simulations  
 12 progressively diverged from the measurements towards crop maturity. For cereals, this might  
 13 be due to the fact that reallocation of assimilates from leaves and stem to storage organs was  
 14 not implemented in AgroC (Spitters et al., 1989).

15 In Merzenhausen, LAI and biomass measurements were only conducted at harvest in 2012  
 16 and during the entire growing season in 2013 (both winter wheat). For model calibration over  
 17 the complete simulation period, measurements of plant height were therefore considered. A  
 18 relation between LAI and plant height was determined for 2013. Plant height showed distinct  
 19 differences between 2012 and 2013. In 2013, a smaller height and consequently a lower LAI  
 20 and dry matter allocation were observed. This could not be reproduced by the model only  
 21 based on differences in meteorological conditions in these two years. Winter wheat varieties  
 22 and management differed between the two cultivation periods, and according to Spitters et al.  
 23 (1989), plant parameters can vary substantially between species. In addition, it needs to be  
 24 considered that in spring of 2013 pronounced dry conditions came to pass. Even though water  
 25 stress was explicitly accounted for in AgroC, irreversible damages (e.g., by heat stress) of

1 plant tissue might have caused a reduced growth beyond the water stress period. Furthermore,  
2 the root system may have preferably been expanded relative to the shoots due to the water  
3 deficit. These effects were not directly considered in AgroC, and could only be captured by  
4 different parameterizations. Therefore, we ran AgroC with crop parameter sets for winter  
5 wheat that differed between the two cultivation periods.

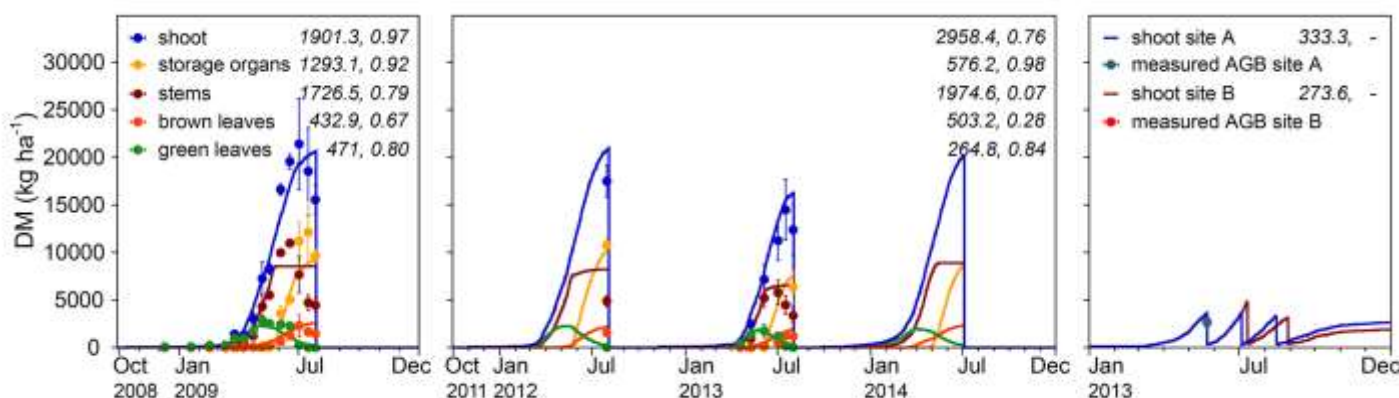
6 The Rollesbroich grassland site was covered by snow until the beginning of April 2013, thus  
7 plant growth was delayed. The model was fitted to the plant development and growth on  
8 parcel A. For the simulation of parcel B, only the dates of mowing were adjusted. This  
9 resulted in an adequate simulation for LAI and dry matter allocation of both grassland parcels  
10 (Fig. 4, 5).

11 At the day of harvest, the simulations for Selhausen and Merzenhausen resulted in mean  
12 root/shoot dry matter ratios of 0.08 and 0.16, respectively. Bolinder et al. (1997, 2002)  
13 determined root/shoot ratios between 0.13 and 0.20 for winter wheat. Compared to this, the  
14 simulated root/shoot ratio for Selhausen was rather low. However, observations of  
15 rhizospheric respiration at this test site (Fig. 6) confirmed the estimated partitioning of  
16 assimilates between shoot and roots. For the Rollesbroich grassland site, the mean root/shoot  
17 ratio was 0.58. This corresponds well with López et al. (2013), who reported a root/shoot ratio  
18 of 0.56 for *Lolium perenne*.



*Fig. 4:*

Observed (dots; error bars: standard deviation) and simulated (lines) leaf area index (LAI) in Selhausen (left), Merzenhausen (middle), and Rollesbroich (right). For the two cropped fields green and brown LAI were measured and simulated. Root mean square error (RMSE) and model efficiency (ME) (in this order) are given for each quantity and location.



*Fig. 5:*

Observed (dots; error bars: standard deviation) and simulated (lines) dry matter (DM) in Selhausen (left), Merzenhausen (middle), and Rollesbroich (right; AGB: above-ground biomass). Root mean square error (RMSE) and model efficiency (ME) (in this order) are given for each quantity and location.



## 1 *Soil Respiration*

2 Magnitude and dynamics of soil CO<sub>2</sub> efflux were captured adequately by AgroC, as shown by  
 3 *ME* values larger than 0.58, *RMSE* values lower than 45.4 mol ha<sup>-1</sup> h<sup>-1</sup>, and an *r* larger than  
 4 0.77 across all sites. For the Selhausen site, observations of efflux due to heterotrophic  
 5 respiration were available separately (Prolingheuer et al., 2014). Therefore, Figure 6 shows  
 6 not only modeled total respiration, but also the simulated partitioning in root and rhizosphere  
 7 respiration and heterotrophic respiration. Since this partitioning is available only for the  
 8 production terms but not for efflux at the surface, the errors reported in Figure 6 differ slightly  
 9 from those presented above. Parameters of the reduction functions for heterotrophic CO<sub>2</sub>  
 10 production in the soil profile were estimated inversely. The start parameter for the reference  
 11 temperature was set to the annual mean temperature at each site as suggested by Coleman and  
 12 Jenkinson (2008). In the optimization process, all reference temperatures were decreased, thus  
 13 CO<sub>2</sub> production was increased at any temperature. As reported by Bauer et al. (2012) and  
 14 Moyano et al. (2012), the approach after Skopp et al. (1990) provided the best results for the  
 15 response of CO<sub>2</sub> production to soil moisture. Therefore, the two control parameters of this  
 16 response function were calibrated. The estimated optimal water content (maximum of  
 17 reduction function curve) was 0.41, 0.29, and 0.28 cm<sup>3</sup> cm<sup>-3</sup> in Selhausen, Merzenhausen, and  
 18 Rollesbroich, respectively. The optimum water contents were very close to the mean soil  
 19 water content of each simulation (0.38, 0.29, and 0.32 cm<sup>3</sup> cm<sup>-3</sup>, respectively).  
 20 As shown in Figure 6, CO<sub>2</sub> production at the grassland site was higher than at the cropped  
 21 sites, which is attributed to the higher soil organic carbon content (Tab. A.1) and an extensive  
 22 perennial root system. However, the magnitude of the simulated rhizospheric respiration  
 23 turned out to be quite similar for all sites, even though the grassland accumulates root biomass  
 24 over the years. The root/shoot ratios reported above showed that the below-ground  
 25 translocation of assimilated carbon was much higher for grassland than for cereal crops.

Hence, the relative fraction of assimilates partitioned to the root system is larger in grasslands (Kuzyakov and Domanski, 2000). Considering the same growth period, the absolute translocation of carbon is the same for both ecosystems; whilst cereals have a higher productivity per unit area and time, their carbon assimilation is restricted to a shorter growth period compared to grasslands. Further, grasslands are not ploughed, so they are potentially a larger sink for atmospheric carbon (Kuzyakov and Domanski, 2000).

An extensive peak of soil CO<sub>2</sub> emission was simulated right after harvest of the cereals, because a large amount of fresh plant material was added to the carbon pools of the soil. Unfortunately, no chamber-based  $R_{\text{soil}}$  observations were available for those critical time periods to validate these model predictions.

The estimated mean annual ratio between rhizospheric respiration and total  $R_{\text{soil}}$  was 0.12 for Selhausen, 0.21 for Merzenhausen, and 0.34 for Rollesbroich. Wang and Fang (2009) analyzed 36 grassland sites and reported a corresponding average ratio of 0.36, which agrees well with results for our grassland site in Rollesbroich. For winter wheat, Moureaux et al. (2008) obtained a ratio between below-ground respiration by autotrophs and total  $R_{\text{soil}}$  of 0.56 for the vegetation period only. Suleau et al. (2011) found ratios between 0.40 and 0.48 using root exclusion experiments. The simulated ratios for the vegetation period were 0.18 for Selhausen and between 0.33 and 0.38 for Merzenhausen. It seems that the simulated fraction of rhizospheric respiration in Selhausen is too low compared to previous studies. However, these values were confirmed by measurements from root exclusion experiments at this site (Prolingheuer et al., 2014). Subke et al. (2006) compared numerous respiration ratios derived by various methods from several studies, and report that the heterotrophic source term may be overestimated by root exclusion, because of increased dead root biomass (for experiments conducted within perennial vegetation), a change of irradiation, and a decreased water uptake by roots. In our study, those error sources were mostly excluded, due to installation of the

1 exclusion rings before cereal growth, a small ring size that enables representative growth and  
2 shading around/above the measurement points, and the correction for the soil moisture effects  
3 (Prolingheuer et al., 2014).  
4 For Rollesbroich, measurements of soil CO<sub>2</sub> concentration in different depths were available,  
5 which allowed calibration of the CO<sub>2</sub> flux through the soil. The approach after Kristensen et  
6 al. (2010), which additionally accounts for diffusion through fractures and macropores,  
7 provided the best results with a *ME* of 0.44 (Fig. 7).

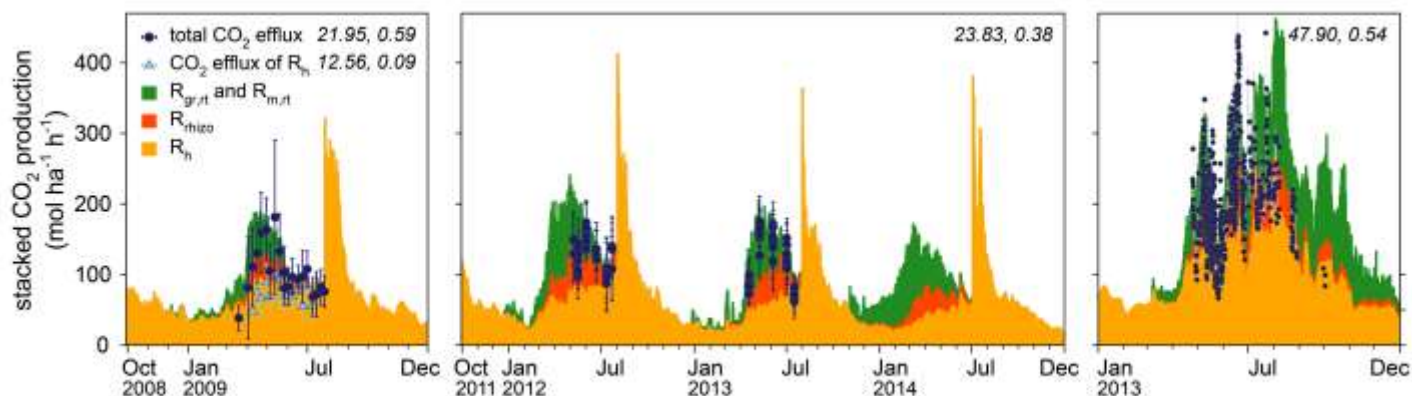


Fig. 6:

Observed (dots; error bars: standard deviation) CO<sub>2</sub> efflux at soil surface and simulated stacked CO<sub>2</sub> production in soil profile (areas) for several source terms (green: growth and maintenance respiration by roots ( $R_{gr,rt}$ ,  $R_{m,rt}$ ); orange: respiration in rhizosphere ( $R_{rhizo}$ ) due to root exudates and root decay; yellow: respiration by heterotrophs ( $R_h$ )) in Selhausen (left), Merzenhausen (middle), and Rollesbroich (parcel A, right). Root mean square error (RMSE) and model efficiency (ME) (in this order) are given for each location.

10

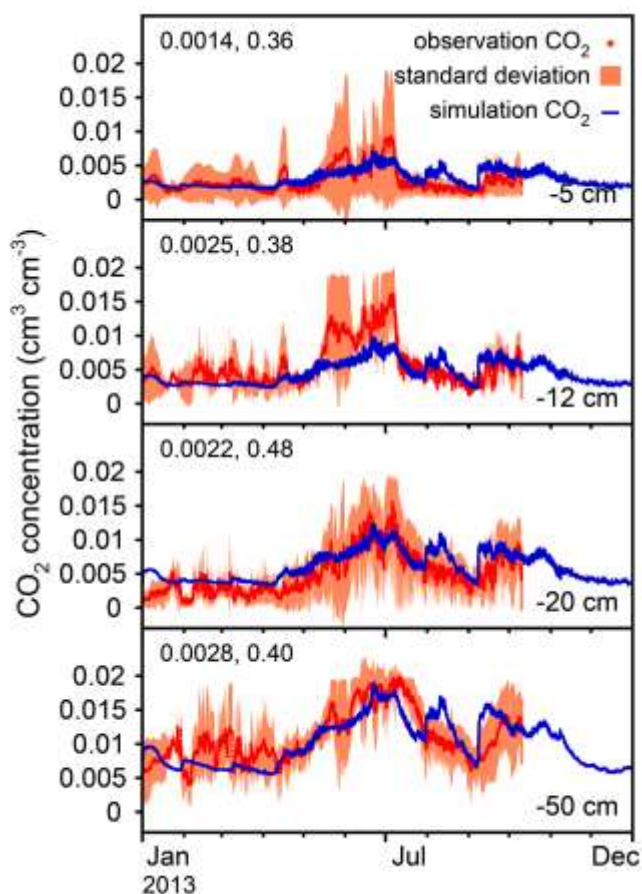


Fig. 7:

Observed (dots; orange area: standard deviation) and simulated (lines) soil CO<sub>2</sub> concentration at various depths in Rollesbroich. Root mean square error (RMSE) and model efficiency (ME) (in this order) are given for each soil depth.

# 1 *Net Ecosystem Exchange*

2 After calibrating soil water flux, plant development, and CO<sub>2</sub> flux, we compared the NEE  
 3 simulations to the EC measurements at each test site. At this point, NEE measurements were  
 4 not used to calibrate the model. Figure 8 and 9 show the AgroC estimates in comparison to  
 5 the NEE flux measurements. With a *RMSE* between 113 and 128 mol ha<sup>-1</sup> h<sup>-1</sup>, a *ME* between  
 6 0.78 and 0.83, and an *r* between 0.91 and 0.96, AgroC performed reasonably well at all three  
 7 test sites. However, some discrepancies could also be observed. As already discussed for R<sub>soil</sub>,  
 8 the estimated peaks of R<sub>soil</sub> and corresponding NEE after harvest were also not observed in  
 9 the EC measurements (Fig. 8). Fluxes from adjacent and cropped fields could have distorted  
 10 the measurements of the area of interest (e.g., Massman and Lee, 2002). In Merzenhausen in  
 11 autumn 2012, negative CO<sub>2</sub> fluxes were measured even though the crop was harvested. This  
 12 was not captured by the AgroC model, because it was assumed that the field was bare fallow.  
 13 In reality, weeds and wheat emerged again during this post-harvest period and assimilated  
 14 CO<sub>2</sub> until ploughing (cf., Sus et al., 2010).

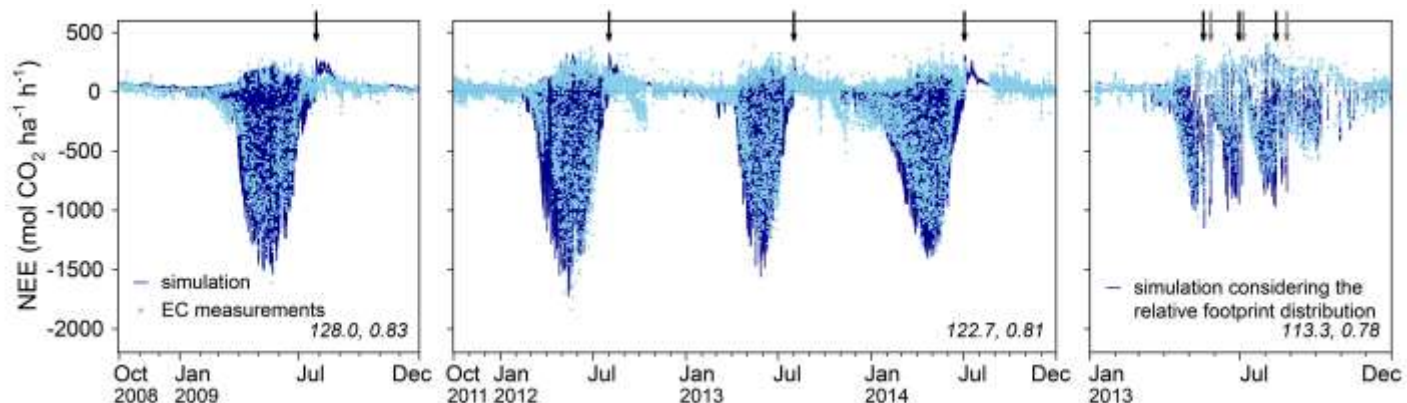
15 At the Rollesbroich site, the EC tower was located at the border between two differently  
 16 managed grassland parcels, so that the contribution of CO<sub>2</sub> fluxes originating from each of the  
 17 two parcels varied according to the flux footprint (Kormann and Meixner, 2001; Mauder et  
 18 al., 2013; Post et al., 2015). For the validation, two AgroC model runs were made for  
 19 grassland parcels A and B. The two NEE estimates were weighted according to the relative  
 20 fraction of the footprint within each parcel, and subsequently compared to the observations.  
 21 Consequently, simulated fluxes could only be attained for time steps at which measurements  
 22 and thus information about the footprint distribution were available. The consideration of the  
 23 footprint distribution improved the performance of the NEE simulations significantly  
 24 compared to a single model run. This was especially true for time periods between two  
 25 mowing events, since parcel B was always mown a few days later than parcel A. Generally,

1 AgroC reproduced the dynamics of the grassland NEE including the effect of mowing and  
2 regrowth. At the time of mowing, leaf area was reduced substantially, canopy photosynthesis  
3 decreased, and the site temporarily turned from a CO<sub>2</sub> sink to a CO<sub>2</sub> source. From the first to  
4 the third mowing, peak assimilation declined consistently. This has previously also been  
5 reported for other grassland sites (Schmitt et al., 2010; Wohlfahrt et al., 2008).

6 The ratios between the annual sum of TER and GPP were 0.79 for Selhausen, between 0.67  
7 and 0.75 for Merzenhausen, and 1.06 for Rollesbroich. The ratios for the growing period only  
8 were 0.64 for Selhausen and between 0.52 and 0.62 for Merzenhausen. The value higher than  
9 1 for Rollesbroich indicates that this site was a CO<sub>2</sub> source in 2013. The annual ratios  
10 between respiration by heterotrophs and TER varied between 0.51 and 0.58 (ratios for  
11 growing period: 0.35 - 0.48). Moureaux et al. (2008) and Suleau et al. (2011) report TER/GPP  
12 ratios between 0.49 and 0.66 for cereals, and R<sub>h</sub>/TER ratios between 0.2 and 0.24, again only  
13 considering the plant growth phase. Our simulations generally agree well with these values,  
14 although the heterotrophic component appears to be larger in this study. Again, this reflects  
15 the lower contribution of rhizospheric respiration as already discussed above.

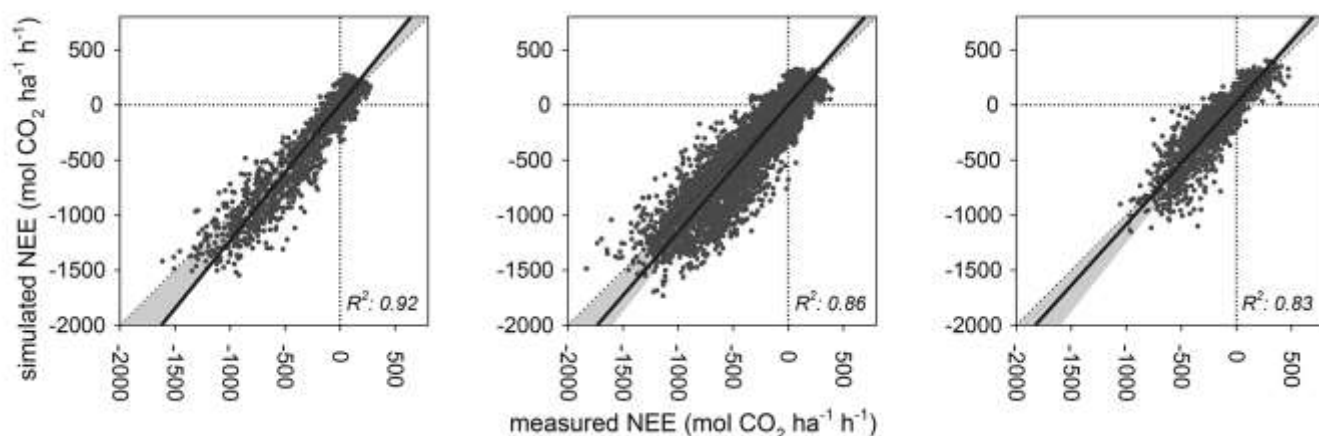
16 The 1:1 plots between observed and simulated NEE (Fig. 9) show that on average AgroC  
17 overestimated the CO<sub>2</sub> fluxes by less than 20%, since the regression lines fall within the grey  
18 area. Turbulence fluxes can be systematically underestimated by EC measurements, and  
19 energy balance closure gaps of this magnitude have previously been reported (Eder et al.,  
20 2015; Schmidt et al., 2012; Twine et al., 2000). Therefore, underestimation of CO<sub>2</sub> fluxes can  
21 be expected (Ingwersen et al., 2015; Massman and Lee, 2002; Mauder et al., 2013). This  
22 inability to close the surface energy balance, the various approaches to correct for the balance  
23 gaps, uncertainties due to instrumentation, and differing data-processing strategies complicate  
24 cross-site and long-term comparisons of NEE (Massman and Lee, 2002; Mauder et al., 2013;  
25 Schmidt et al., 2012; Twine et al., 2000).

1 Wattenbach et al. (2010) compared the efficiency of four models to simulate NEE, and  
2 reported *ME* values between -0.15 and 0.87. The *ME* values for AgroC for the three sites  
3 compare favorably with this wide range (0.78 - 0.83). Wattenbach et al. (2010) also reported  
4 more substantial discrepancies between observations and simulations for positive NEE fluxes.  
5 Such an underestimation of positive NEE fluxes was also observed in this study, but to a  
6 much smaller extent, which is very likely a result of our more advanced approach towards the  
7 simulation of CO<sub>2</sub> fluxes and the calibration of R<sub>soil</sub> with chamber measurements.



*Fig. 8:*

Observed (dots) and simulated (lines) net ecosystem exchange (NEE) in Selhausen (left; EC: eddy covariance), Merzenhausen (middle), and Rollesbroich (right). In Rollesbroich NEE was simulated for each grassland (parcel A and B) and then allocated with the relative fraction of the footprint on each grassland. Arrows indicate dates of harvest or mowing (black: parcel A; grey: parcel B), respectively. Root mean square error (RMSE) and model efficiency (ME) (in this order) are given for each location.



*Fig. 9:*

Observed and simulated net ecosystem exchange (NEE) with regression line (black) in Selhausen (left), Merzenhausen (middle), and Rollesbroich (right). In Rollesbroich NEE was simulated for each grassland (parcel A and B) and then weighted according to the relative fraction of the footprint. A potential NEE gap of up to 20% in the measurements is indicated by the grey area. Coefficient of determination ( $R^2$ ) is given for each location.



### 3.2. Calibration with NEE Data

Due to calibration the *RMSE* of instantaneous NEE was reduced by up to 43%, and *Bias* was severely decreased (Fig. 10). Depending on the optimization strategy, the cumulative NEE over the simulation period differed strongly (Fig. 10, B.3). The calibration based on the instantaneous NEE data ( $NEE_{inst}$ ) yielded the best results in terms of *RMSE*, *ME*, and *r* at all sites, because the reduction of the squared residual error in NEE was the only criterion. *Bias* was the lowest in the  $NEE_{BSc}$  approach with and without inclusion of  $R_{soil}$  data because the *Bias* was now part of the objective function. Apart from that, model performance and NEE prediction by the  $NEE_{BSc} (+ R_{soil})$  approach were very similar to  $NEE_{inst} (+ R_{soil})$ . The  $NEE_{Cum}$  and  $NEE_{Log} + R_{soil}$  approaches resulted in the poorest model performances at each study site. In almost all cases, model performance for NEE slightly deteriorated when  $R_{soil}$  measurements were included in the optimization process due to trade-offs between fitting multiple objective functions, with the exception of the approach that considered  $NEE_{Cum} + R_{soil}$  (Fig. 10).

Figure 11 shows reduced major axis regression (Webster, 1997) for measured and simulated day- and nighttime (nighttime hours with global radiation  $< 20 \text{ W m}^{-2}$  after Reichstein et al., 2005) NEE fluxes for the test site Selhausen. The corresponding figures for Merzenhausen and Rollesbroich are given in the appendix (Fig. B.1, B.2). Compared to the NEE runs obtained without calibration (Fig. 9), the calibrated daytime fluxes were generally closer to the 1:1 line and tended to only slightly underestimate daytime NEE fluxes as indicated by regression slopes slightly lower than 1. In general, nighttime NEE fluxes (dominated by respiratory fluxes) were better captured by the approaches that used an objective function including  $R_{soil}$  data, irrespective of the error weighting in the objective function or the transformation of the raw NEE data. Including  $R_{soil}$  data in the calibration clearly improved the simulation of diurnal and annual dynamics of the measured  $R_{soil}$ . The approaches only

considering NEE measurements did not reproduce those dynamics (not shown). Even with the inclusion of  $R_{soil}$  data, nighttime NEE was still underestimated as indicated by regression slopes between 0.75 and 0.85.

In Figure 10 (bottom right panel) and in the appendix (Fig. B.3), cumulative NEE over the corresponding simulation period (further on, only referred to as “cumulative NEE”) is shown for all optimization strategies, for the simulations without calibration, and for the gap-filling method by Reichstein et al. (2005). For this comparison, cumulative NEE estimated with AgroC was also calculated in a “gap-filling mode”, keeping the EC measurements and only filling the gaps with AgroC results. The cumulative NEE varied between -462 and -243 g C m<sup>-2</sup> in Selhausen, -1429 and -1180 g C m<sup>-2</sup> in Merzenhausen, and -541 and -5 g C m<sup>-2</sup> in Rollesbroich. Cumulative NEE was mostly lower for the calibrated model runs than for the uncalibrated simulation. For all sites, the  $NEE_{Cum}$  or  $NEE_{Log}$  approach with and without  $R_{soil}$  measurements resulted in the lowest cumulative NEE. The  $NEE_{inst} + R_{soil}$  approach resulted in the highest NEE, except for the Rollesbroich site. Generally, cumulative NEE of approaches including  $R_{soil}$  data in the objective function showed better agreement with the gap-filling method after Reichstein et al. (2005) than the approaches that did not consider  $R_{soil}$  measurements (Fig. 10).

Neglecting carbon removal due to harvest, the simulations suggest that all sites are CO<sub>2</sub> sinks, except for the simulation without calibration to NEE in Rollesbroich, which showed a very small positive annual NEE. Pastures are usually considered to be sinks for atmospheric CO<sub>2</sub> (Kuziyakov and Domanski, 2000). Soussana et al. (2007) estimated an average annual carbon budget of  $-247 \pm 67$  g C m<sup>-2</sup> and a net biome productivity (= NEE minus carbon loss due to disturbances, such as harvest) of  $-104 \pm 73$  g C m<sup>-2</sup> for nine grasslands in Europe. Wohlfahrt et al. (2008) reported alternating positive and negative annual NEE for one grassland (gap-filled EC measurements), varying between  $-42$  g C m<sup>-2</sup> a<sup>-1</sup> and  $69$  g C m<sup>-2</sup> a<sup>-1</sup>, and concluded

1 that meteorological variations or differing biotic responses could easily lead to a positive  
2 carbon balance in some years. Also, the large amount of carbon stored in grassland soils  
3 (Tab. A.1) can easily cause large respiratory fluxes that exceed plant carbon uptake. For  
4 Selhausen, estimated NEE matches cumulative values reported by Schmidt et al. (2012) and  
5 Wattenbach et al. (2010). Anthoni et al. (2004) found annual NEE in a range from -185  
6 to  $-245 \text{ g C m}^{-2}$  for a winter wheat field in Germany in 2001, which is in good agreement with  
7 our findings.

8 Since the true cumulative NEE is unknown due to measurement gaps, modelling can provide  
9 valuable information about the carbon balance. Although the best calibration approach that  
10 provides the 'true' cumulative NEE cannot be determined at this point. Our results suggest  
11 that the cumulative NEE obtained from the calibrated model runs is more realistic than the  
12 cumulative NEE obtained with a model run not calibrated to NEE. The well-established gap-  
13 filling method after Reichstein et al. (2005) and AgroC produced somewhat different carbon  
14 balances, although NEE was derived from the same weather data. Especially after harvest or  
15 mowing, AgroC provided more reasonable predictions because it considers the changes in  
16 crop characteristics that directly influence GPP. Nevertheless, a better representation of  
17 respiration processes is still required, because even after calibration with EC and chamber  
18 measurements the respiration by heterotrophs and autotrophs was still underestimated. This  
19 bias in respiration may indicate a wrong process representation in the model, errors in model  
20 parameterization, or may also be related to a disparity in the measurement footprint between  
21 chamber and EC measurements (Richardson et al., 2010). Obviously, an underestimation of  
22 respiratory fluxes will shift NEE to more negative values, as observed for the simulation  
23 results in Figure 10.

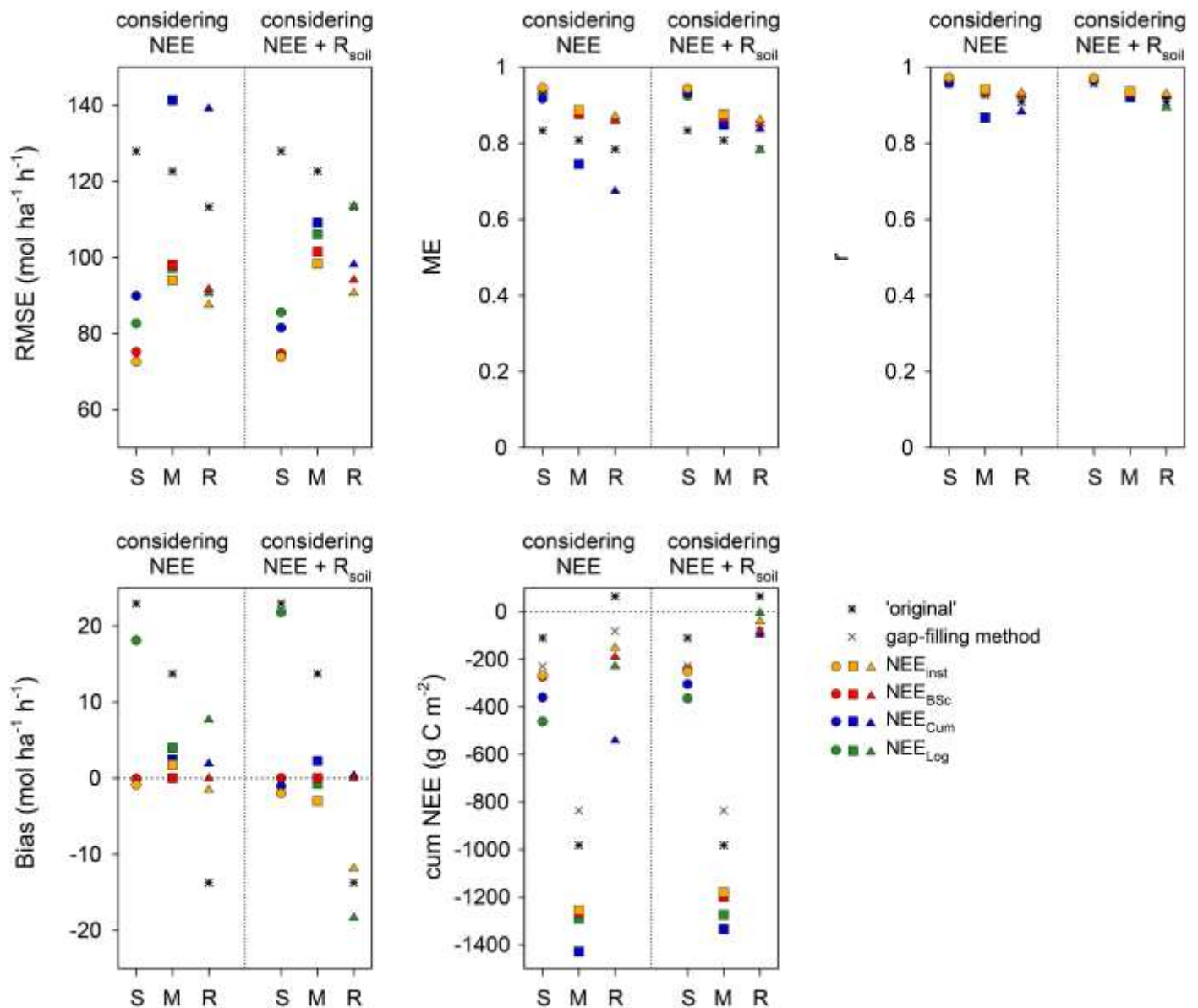
24 The cumulative NEE obtained after calibration with EC measurements was sensitive to the  
25 definition of the objective function and the data-transformation. As expected, explicit

consideration of *Bias* in the objective function reduced the *Bias* substantially (Fig. 10), with the  $NEE_{BSc}$  approach being most effective. The  $NEE_{Cum}$  approach often led to overestimation of negative and underestimation of positive fluxes (Fig. 10, 11, B.1, B.2). The use of cumulative data is known to enhance systematic errors and reduce noise (Hess and Schmidt, 1995; Mandel, 1957), and might not provide statistically valid information about associated errors and results if non-random auto-correlated residuals prevail. Compared to using the *Bias* as a criterion, it gives more weight to early observations that affect all succeeding cumulative values in the simulation period.

High-quality (hourly) EC measurements obtained after data processing usually consist of a large number of large negative fluxes during daytime and a smaller number of small positive nighttime fluxes, the latter being underrepresented. During calibration, the negative fluxes will on average have a higher weight, since they are more frequent and larger than positive fluxes. Therefore, a log-transformation of the NEE data could partly compensate for this, and provide more equal weighting. However, our results suggest the effect of this transformation on the performance of the calibration was weak. The slope of the regression between observed and simulated positive NEE was just slightly closer to 1 for the  $NEE_{Log} (+ R_{soil})$  approach (Fig. 11, B.1, B.2).

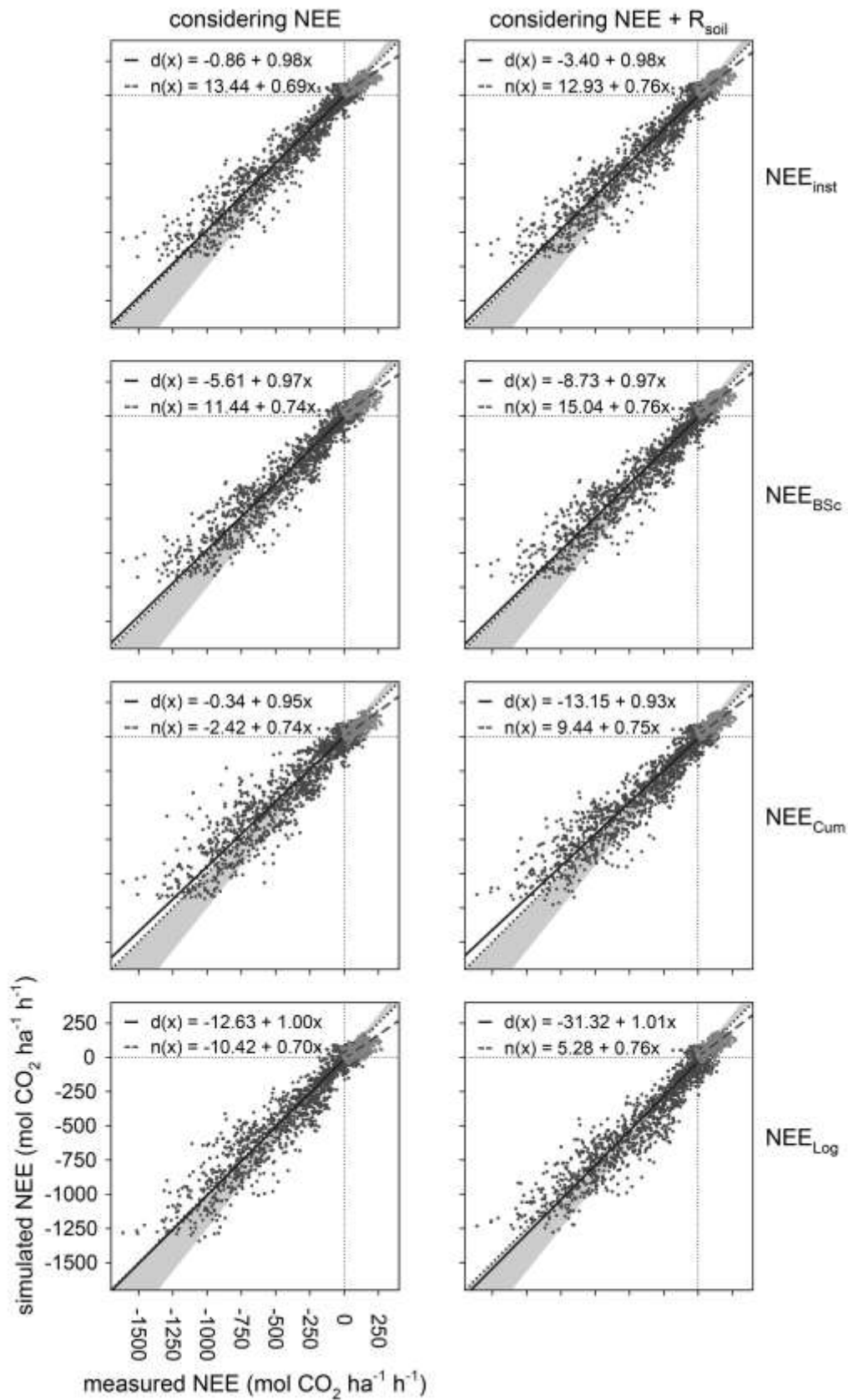
The model performance for small positive fluxes improved strongly when considering  $R_{soil}$  measurements as an additional data source (Fig. 11, B.1, B.2). Similar findings were reported by Richardson et al. (2010), Wang et al. (2009), and Yuan et al. (2012). Williams et al. (2009) stated that usage of multiple data streams in an inverse estimation lessens the criticalness of biases and internal inconsistencies in each data stream. Including  $R_{soil}$  measurements in the optimization process notably reduced the bias observed in the simulation of nighttime NEE more than any of the modifications of the objective function or the use of data-transformation.

1 The  $NEE_{inst} + R_{soil}$  approach provided the best results regarding both day- and nighttime  
2 fluxes at all three test sites. On average, model bias was one of the lowest for this  
3 optimization strategy at all sites. Even though overall model performance of the eight  
4 calibration approaches differed only marginally, resulting cumulative NEE diverged strongly.  
5 Considering additional data sources such as biomass measurements should help to further  
6 decrease the uncertainty of the cumulative NEE estimation (Richardson et al., 2010).



*Fig. 10:*

Root mean square error (RMSE), model efficiency (ME), Pearson product-moment correlation coefficient (r), Bias, and cumulated net ecosystem exchange (cum NEE) over simulation time period, calculated in “gap-filling mode”, for each optimization strategy, for the simulation without calibration to NEE (‘original’), and for the gap-filling method after Reichstein et al. (2005) (gap-filling method) at all three study sites (S: Selhausen; M: Merzenhausen; R: Rollesbroich). For description of optimization strategies see text.



1 *Fig. 11:*  
2 Correlations between observed and simulated net ecosystem exchange (NEE) for all  
3 optimization strategies at test site Selhausen. Reduced major axis regression was derived for  
4 each strategy distinguished between day- (d) and nighttime (n) CO<sub>2</sub> fluxes, whereat nighttime  
5 was designated to a measured global radiation lower than 20 W m<sup>-2</sup>. For description of  
6 optimization strategies see text.

#### 4. Conclusions

The present study demonstrates that a crop growth module coupled to a model of soil CO<sub>2</sub> production, soil water and heat flux can be used to simulate hourly NEE in agricultural systems. After calibrating the model for soil moisture, crop development, and  $R_{soil}$ , the simulation of hourly NEE agreed well to EC measurements. For further validation, the application of AgroC to cropping systems in different European climate regions would be interesting.

An additional calibration based on EC measurements further improved the model in terms of the performance criteria. Even more importantly, systematic errors between EC data and model were reduced. However, the various calibration approaches reveal that particularly the cumulative NEE over the entire simulation period is rather strongly affected by the choice of the objective criterion. Based on the evaluation of different optimization strategies, we recommend the use of the *RMSE* and non-transformed instantaneous EC-derived fluxes in combination with  $R_{soil}$  measurements (if available) by equally weighted errors. Our results indicate that inversely estimated and gap-filled cumulative NEE is associated with considerable uncertainty, which can be decreased when  $R_{soil}$  measurements are included in the optimization process. At the same time, inclusion of  $R_{soil}$  also provided a substantial reduction of bias in the simulation of the respiratory fluxes.

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# APPENDIX

## Appendix A: The AgroC Model

### *Hourly Time Step*

The SOILCO2/RothC model has a flexible time stepping scheme, however the original SUCROS model explicitly runs at a daily time step. Since NEE typically shows distinct diurnal variations, the SUCROS code was adapted to work with an hourly time step. Only the calculation of development stage DVS (-) still relies on the original approach based on the effective temperature sum. In the SUCROS model, daily total gross assimilation is obtained by three-point Gauss integration of the instantaneous assimilation rates per unit leaf area over the daylight period. This integration was omitted in the AgroC model with an hourly time step. Hourly gross assimilation is computed from the hourly average inputs of global radiation and mean temperature using the same approach that was used for the instantaneous assimilation rate in the original code. Major changes were required for the estimation of the photosynthetic active radiation (PAR) flux at the top of the canopy. In SUCROS, instantaneous PAR ( $\text{J [L]}^{-2} [\text{T}]^{-1}$ ) is estimated from the sine of solar inclination  $\sin B$  (-) and the daily integral of  $\sin B$  including a correction of lower atmospheric transmittance at lower solar elevation  $d\sin BE$  ( $\text{s d}^{-1}$ ). The integral daily value  $d\sin BE$  is approximated and  $\sin B$  is estimated for the day of the year in dependence of the geographic position. In AgroC, the hourly integral of the sine of solar inclination  $d\sin B$  ( $\text{s h}^{-1}$ ) is now calculated using the trapezoidal rule according to:

$$d\sin B = 0.5 \left( \sin B_{t-1} + (\sin(\delta) \sin(\varphi) + \cos(\delta) \cos(\omega) \cos(\varphi)) \right) t_s \quad (\text{A.1})$$

where instantaneous  $\sin B_{t-1}$  ( $= \sin(\delta) \sin(\varphi) + \cos(\delta) \cos(\omega) \cos(\varphi)$ ) is the sine of solar elevation of the previous hour,  $\delta$  ( $^\circ$ ) is the sun declination angle,  $\varphi$  ( $^\circ$ ) is the geographic latitude,  $\omega$  ( $^\circ$ ) is the hour angle, and  $t_s$  (s) is the number of seconds with astronomically possible solar radiation within one hour (3600 during day, 0 during night, and a value in between for the two hours that include sunrise and sunset). The value of  $d\sin BE$  is then estimated as:

$$d\sin BE = \sin \left( \arcsin(0.5 (\sin B_{t-1} + \sin B)) + 0.4 (0.5 (\sin B_{t-1} + \sin B)) \right) t_s \quad (\text{A.2})$$

where 0.4 is the regression coefficient between transmission and solar angle (Supit et al., 1994).

### Water Fluxes

The coupling between SOILCO2 and SUCROS involves two hydrological processes: rainfall interception and root water uptake. Interception loss is estimated according to the single-big-leaf concept (Rutter et al., 1971). The canopy interception storage capacity  $S_i$  ([L]) was assumed to be proportional to the total leaf area index  $LAI$  ( $[L^2 L^{-2}]$ ). Water is removed from the interception storage by evaporation  $E_i$  ( $[L T^{-1}]$ ):

$$E_i = (ET_{p,crop} - E_p) \frac{C_i}{S_i} \quad (\text{A.3})$$

where  $C_i$  ([L]) represents the interception storage at a certain time step,  $ET_{p,crop}$  ( $[L T^{-1}]$ ) is the potential crop evapotranspiration, and  $E_p$  ( $[L T^{-1}]$ ) is the potential soil evaporation. The amount of interception  $N_i$  ( $[L T^{-1}]$ ) is then estimated according to:



$$N_i = \begin{cases} 0 & N_0 = 0 \\ S_i - C_i & \text{for } S_i - C_i < N_0 \\ N_0 & S_i - C_i > N_0 \end{cases} \quad (\text{A.4})$$

2

3 where  $N_0$  ( $[L\ T^{-1}]$ ) represents precipitation. The amount of precipitation entering the soil  $N_p$   
 4 ( $[L\ T^{-1}]$ ) is calculated as the difference between  $N_0$  and  $N_i$ .

5 In SUCROS,  $ET_{p,crop}$  is computed by scaling the potential grass reference evapotranspiration  
 6 (Penman-Monteith approach; Allen et al., 1998) with the dimensionless crop conversion  
 7 factor  $K_c$ . On the basis of Beer's law,  $ET_{p,crop}$  is split into potential soil evaporation  $E_p$   
 8 ( $[L\ T^{-1}]$ ) and potential transpiration  $T_p$  ( $[L\ T^{-1}]$ ) in dependence of the  $LAI$ :

9

$$E_p = ET_{p,crop} \exp(-0.6 \cdot LAI) \quad (\text{A.5})$$

$$T_p = ET_{p,crop} - E_p - E_i \quad (\text{A.6})$$

12

13 The potential soil evaporation is passed to SOILCO2, where it is used to prescribe the  
 14 potential upward water flux as upper boundary condition. Potential transpiration is distributed  
 15 over soil depth according to the relative root density distribution to provide the potential sink  
 16 term for root water uptake. The depth-specific actual root water uptake is computed by scaling  
 17 the potential root water uptake with reduction factor  $\alpha$  (-) in dependence of soil pressure head  
 18  $h$  ( $[L]$ ) following the approach of Feddes et al. (1978):

19

$$\alpha(h) = \begin{cases} \frac{h_0 - h}{h_0 - h_1} & h_0 \leq h \leq h_1 \\ 1 & \text{for } h_1 \leq h \leq h_2 \\ 10^{\frac{h_2 - h}{h_3}} & h_2 \leq h \leq h_3 \end{cases} \quad (\text{A.7})$$

21

22 where  $h_0$ ,  $h_1$ ,  $h_2$ , and  $h_3$  ( $[L]$ ) are prescribed threshold pressure heads (Vanclooster et al.,  
 23 1995), which are plant dependent (Tab. A.2). Integration of the actual root water uptake over

depth provides the actual transpiration  $T_a$  ( $[L\ T^{-1}]$ ). The reduction of stomatal conductance due to water stress was assumed to correspond to the ratio between actual and potential transpiration  $T_a/T_p$ .

#### Carbon Fluxes

In this study, carbon fluxes from the atmosphere to the ecosystem (downward) are defined as negative fluxes, and upward fluxes are defined as positive. The water stress ratio ( $T_a/T_p$ ) is used to scale gross carbon assimilation and to account for the effect of limited soil water availability on crop activity in terms of gross primary productivity  $GPP$  ( $\text{mol CO}_2 [L]^{-2} [T]^{-1}$ ):

$$GPP = -\frac{G_{phot}}{Mol_{CH_2O}} \cdot \frac{T_a}{T_p} \quad (A.8)$$

where  $G_{phot}$  ( $\text{kg CH}_2\text{O } [L]^{-2} [T]^{-1}$ ) is the glucose equivalent of the total gross assimilation per time step (Spitters et al., 1989), and  $Mol_{CH_2O}$  is the molar mass of  $\text{CH}_2\text{O}$  ( $= 0.030 \text{ kg mol}^{-1}$ ).

The net primary productivity  $NPP$  ( $\text{mol CO}_2 [L]^{-2} [T]^{-1}$ ) is defined as:

$$NPP = GPP + R_{gr} + R_m \quad (A.9)$$

where  $R_{gr}$  ( $\text{mol CO}_2 [L]^{-2} [T]^{-1}$ ) is the total growth respiration, and  $R_m$  ( $\text{mol CO}_2 [L]^{-2} [T]^{-1}$ ) is the maintenance respiration. Net ecosystem exchange  $NEE$  ( $\text{mol CO}_2 [L]^{-2} [T]^{-1}$ ) is computed as:

$$NEE = NPP + R_h \quad (A.10)$$

where  $R_h$  ( $\text{mol CO}_2 [\text{L}]^{-2} [\text{T}]^{-1}$ ) is the depth-integral of the heterotrophic  $\text{CO}_2$  source term provided by the RothC module.

#### *Maintenance and Growth Respiration*

In a first step, the total maintenance respiration demand at  $25^\circ\text{C}$   $R_{m,r}$  ( $\text{kg CH}_2\text{O} [\text{L}]^{-2} [\text{T}]^{-1}$ ) is computed as a glucose equivalent according to:

$$R_{m,r} = \sum_{o=1}^4 f_{m,o} W_o f_t \quad (\text{A.11})$$

where  $f_{m,o}$  ( $\text{kg CH}_2\text{O kg}^{-1} \text{ DM} [\text{T}]^{-1}$ ) is the maintenance coefficient with index  $o$  looping over the four plant organs leaves, stems, roots, and storage organs with coefficients of 0.03, 0.015, 0.015, and 0.01, respectively (Spitters et al., 1989),  $W_o$  ( $\text{kg DM} [\text{L}]^{-2}$ ) is the respective organ dry weight, and  $f_t$  (-) is a time conversion factor accounting for the use of an hourly or daily time step. In a second step,  $R_{m,r}$  is corrected for temperature to estimate total maintenance respiration  $R_{m,c}$  ( $\text{kg CH}_2\text{O} [\text{L}]^{-2} [\text{T}]^{-1}$ ) as described by Spitters et al. (1989) and converted to  $\text{CO}_2$  equivalent maintenance respiration  $R_m$  ( $\text{mol CO}_2 [\text{L}]^{-2} [\text{T}]^{-1}$ ) by dividing with  $\text{Mol}_{\text{CH}_2\text{O}}$ .

Total growth respiration  $R_{gtot}$  ( $\text{kg CH}_2\text{O} [\text{L}]^{-2} [\text{T}]^{-1}$ ) in glucose equivalents is estimated as:

$$R_{gtot} = \left( G_{phot} \cdot \frac{T_a}{T_p} - R_{m,c} \right) - \Delta W \cdot C_{cont} \cdot \frac{\text{Mol}_{\text{CH}_2\text{O}}}{\text{Mol}_C} \quad (\text{A.12})$$

where  $\Delta W$  ( $\text{kg DM} [\text{L}]^{-2} [\text{T}]^{-1}$ ) is the overall dry matter growth rate,  $C_{cont}$  ( $\text{g C g}^{-1} \text{ DM}$ ) is the conversion factor between carbon and biomass dry matter weight, and  $\text{Mol}_C$  is the molar mass of C ( $= 0.012 \text{ kg mol}^{-1}$ ). Growth respiration for each plant organ  $R_{gr,o}$  ( $\text{mol CO}_2 [\text{L}]^{-2} [\text{T}]^{-1}$ ) is computed from  $R_{gtot}$  according to:

$$R_{gr,o} = \frac{R_{gtot} \cdot f_o}{Mol_{CH_2O}} \quad (A.13)$$

where index  $o$  loops over the four plant organs, and  $f_o$  (-) is the organ-specific partitioning factor. Total growth respiration  $R_{gr}$  (mol CO<sub>2</sub> [L]<sup>-2</sup> [T]<sup>-1</sup>) is finally computed as the sum of all  $R_{gr,o}$ . The sum of maintenance and growth respiration of the roots represents the autotrophic source term of soil CO<sub>2</sub> and is distributed over the soil profile according to the time-variable relative root density distribution.

#### *Root Exudation and Root Decay*

In SUCROS, the daily or hourly glucose assimilation rate  $G_{phot}$  (kg CH<sub>2</sub>O [L]<sup>-2</sup> [T]<sup>-1</sup>) is partitioned in dependence of the DVS into the fraction for the shoot and for the root system to build up biomass. According to labelling experiments performed by Swinnen et al. (1995) for winter wheat, 18.2% of net assimilation is transferred to the roots, 7.1% are used to build up root biomass, and 5.3% are released as young photosynthetate rhizodeposition. This translates into fractions of 0.39 and 0.29 for root biomass build-up and exudates, respectively, relative to net assimilation transferred to the roots. The remaining fraction consists of root respiration and root decay. The relative root exudation factor  $f_{exu}$  (-) thus equals 0.43 (= 0.29 / (0.39 + 0.29)). In AgroC, the root exudation rate  $Rt_{exu}$  (kg C [L]<sup>-2</sup> [T]<sup>-1</sup>) is computed according to this partitioning factor from the dry matter root growth rate  $\Delta W_{rt}$  (kg DM [L]<sup>-2</sup> [T]<sup>-1</sup>):

$$Rt_{exu} = \Delta W_{rt} \cdot f_{rt} \cdot f_{exu} \cdot 0.467 \quad (A.14)$$

where  $f_{rt}$  is the dimensionless partitioning factor for roots, and 0.467 kg C kg<sup>-1</sup> DM is the root-specific dry matter carbon content (Goudriaan et al., 1997). Using this approach, the

simulated root exudation shows diurnal variations due to the dependence on the assimilation rate, as suggested by Hopkins et al. (2013) and Kuzyakov (2006) amongst others. Swinnen et al. (1995) reported that 3.1% of the net assimilation ends up as dead roots. In relation to the 18.2% transferred to the roots, this equals a relative fraction of 0.17. In order to account for this, a root death factor  $f_{dea}$  (-) was introduced. It was assumed that  $f_{dea}$  is lower during the crop juvenile stages than at flowering:

$$f_{dea} = \begin{cases} 0 & DVS < 0.2 \\ \frac{f_{deamax}(DVS-0.2)}{0.5-0.2} & \text{for } 0.2 \leq DVS \leq 0.5 \\ f_{deamax} & DVS > 0.5 \end{cases} \quad (A.15)$$

where  $f_{dea}$  is the root death factor in relation to the total amount of roots, and  $f_{deamax}$  (-) is the maximum value of the root death factor. For winter wheat, a  $f_{deamax}$  of 0.43 was used, which approximately reproduced the cumulative fraction of dead roots of 0.17 of net assimilation determined by Swinnen et al. (1995). The rate of root death in terms of carbon release  $Rt_{dea}$  ( $\text{kg C [L]}^{-2} [\text{T}]^{-1}$ ) is computed as:

$$Rt_{dea} = \Delta W_{rt} \cdot f_{rt} \cdot f_{dea} \cdot 0.467 \quad (A.16)$$

$\Delta W_{rt}$  is reduced according to the loss of root exudates and dead roots. The total amount of root exudates and dead roots is again distributed over depth according to the relative root density profile. The carbon equivalent of the root exudates is transferred to the depth-specific decomposable plant material pool (DPM) of the RothC subroutine because of the expected rapid decomposition of these labile substances by rhizosphere microorganisms. The carbon equivalent of the dead roots is split into the DPM and the resistant plant material (RPM) pool

according to the original RothC partitioning factor for incoming plant material of 0.59 and 0.41 (Coleman and Jenkinson, 2008), respectively.

For winter wheat and barley, harvest residues are also considered. At the time of harvest, root biomass and 25% of stem biomass is added to the DPM and RPM pool up to a user-specified soil depth (i.e. ploughing depth). Figure 1 provides a summary of the carbon cycling in AgroC.

### *Grassland*

The original SUCROS code is not capable of simulating managed grassland, which are characterized by multiple mowing events over the season. Mowing is associated with the transfer of glucose from roots and stubble to the leaves, which allows for a faster compensation of defoliation. The routines implemented in AgroC for the simulation of the above-mentioned processes follow the sink/source approach suggested by Schapendonk et al. (1998) for the grassland productivity model LINGRA.

At prescribed mowing dates, the current green leaf area index  $LAI_g$  is set to a fixed post-mowing leaf area index  $LAI_{post}$  (in this study we set  $LAI_{post} = 0.35$  based on LAI measurements). The ratio between pre-mowing LAI and post-mowing  $LAI_{post}$  is used to compute the respective loss of dry matter biomass:

$$f_{lai} = \frac{LAI_g}{LAI_{post}} \quad (A.17)$$

$$w_{post,i} = \frac{w_{pre,i}}{f_{lai}} \quad (A.18)$$

where  $f_{lai}$  (-) is the pre-/post-mowing LAI ratio,  $w_{pre}$  (kg DM [L]<sup>-2</sup>) is the biomass prior to mowing, and  $w_{post}$  (kg DM [L]<sup>-2</sup>) is the respective biomass after mowing. The index  $i$  loops over leaves, stems, and storage organs/inflorescence. At each mowing event, DVS is also

reset to a prescribed value of  $DVS_{reset} = 0.5$ . In order to simulate the transfer of glucose after defoliation, we implemented a glucose storage that is filled between a  $DVS_{lo}$  of 0.6 and a  $DVS_{hi}$  of 1.0. The rate of glucose storage increase  $\lambda_{s+}$  ( $\text{kg CH}_2\text{O [L]}^{-2} [\text{T}]^{-1}$ ) is computed as a fraction  $f_{stor}$  (-) of global net glucose production:

$$\lambda_{s+} = \left( G_{phot} \cdot \frac{T_a}{T_p} - R_{m,c} \right) \cdot f_{stor} \quad (\text{A.19})$$

The part of global net glucose production ( $= G_{phot} \cdot T_a/T_p - R_{m,c}$ ) available for biomass growth and respiration is reduced accordingly by  $\lambda_{s+}$ . The storage fraction is computed in dependence of DVS:

$$f_{stor} = \begin{cases} 0 & DVS \leq DVS_{lo} \\ \frac{f_{stormax}(DVS - DVS_{lo})}{(DVS_{hi} - DVS_{lo})} & \text{for } DVS_{lo} < DVS < DVS_{hi} \\ f_{stormax} & DVS \geq DVS_{hi} \end{cases} \quad (\text{A.20})$$

where  $f_{stormax}$  (-) is the user-specified maximum storage fraction. Thus, the glucose storage  $S_{stor,t}$  ( $\text{kg CH}_2\text{O [L]}^{-2}$ ) increases by  $\lambda_{s+}$  until a user-defined maximum value of  $S_{stormax}$  ( $\text{kg CH}_2\text{O [L]}^{-2}$ ) is reached. After that,  $S_{stor,t}$  remains constant. After mowing, the dry matter transfer rate  $\lambda_{s-}$  ( $[\text{T}]^{-1}$ ) from  $S_{stor,t}$  to the shoot is estimated as:

$$\lambda_{s-} = \frac{\log(100)}{t_{stor}} \quad (\text{A.21})$$

where  $t_{stor}$  ([T]) is a user-specified time required to reach a value of 1% of the storage at the time of mowing. According to Gonzales et al. (1989) and Prud'homme et al. (1992), the mobilization of carbohydrates in ryegrass is highest during the first 6 days after defoliation

and levels out in a second phase that lasts until 29 days after defoliation. In this study,  $t_{stor}$  was set to 15 days, which results in a  $\lambda_{s-}$  of  $0.31 \text{ d}^{-1}$ . Correspondingly,  $S_{stor,t}$  is reduced down to a limiting value of zero according to:

$$S_{stor,t+1} = S_{stor,t} (1 - \lambda_{s-}) \quad (\text{A.22})$$

The additional dry matter growth rate  $\Delta W_{stor}$  ( $\text{kg DM [L]}^{-2} [\text{T}]^{-1}$ ) resulting from the declining  $S_{stor,t}$  is added to the dry matter growth rate of the shoot  $\Delta W_{sh}$  ( $\text{kg DM [L]}^{-2} [\text{T}]^{-1}$ ), which is the outcome of the photosynthetic activity of the plant. The additional shoot growth rate  $\Delta W_{stor}$  is computed as:

$$\Delta W_{stor} = \frac{S_{stor,t} \lambda_{s-}}{f_{sh} (1.46 f_{lv} + 1.51 f_{st})} \quad (\text{A.23})$$

where  $f_{sh}$ ,  $f_{lv}$ , and  $f_{st}$  are the dimensionless partitioning factors for shoot, leaves, and stems, respectively. The assimilate requirement coefficients of 1.46 and 1.51 in Equation A.23 have a unit of  $\text{kg CH}_2\text{O kg}^{-1} \text{ DM}$  (Spitters et al., 1989).

As suggested by Schapendonk et al. (1998), a mechanism was implemented by which the specific leaf area ( $\text{ha leaf kg}^{-1} \text{ DM}$ ) varies over the season as a function of DVS. Furthermore, a mechanism to distinguish between vegetative and reproductive development of grass was introduced as suggested by Barrett et al. (2004). These two stages of development differ in the productivity of grass and in several major physiological processes that alter the response of the plant to environmental drivers (e.g., Anslow and Green, 1967; Leafe et al., 1974; Parsons, 1988; Robson et al., 1988).



Tab. A.1:

Site-specific soil properties ( $C_{\text{org}}$ : organic carbon content) and inversely estimated hydraulic parameters ( $\theta_r$ : residual water content;  $\theta_s$ : saturated water content;  $\alpha$ : inverse of the bubbling pressure;  $n$ : shape parameter;  $K_s$ : saturated hydraulic conductivity; van Genuchten, 1980).

	soil profile horizons	sand (%)	silt (%)	clay (%)	$C_{\text{org}}$ (%)	$\theta_r$ ( $\text{cm}^3 \text{cm}^{-3}$ )	$\theta_s$ ( $\text{cm}^3 \text{cm}^{-3}$ )	$\alpha$ ( $\text{cm}^{-1}$ )	$n$ (-)	$K_s$ ( $\text{cm h}^{-1}$ )
<b>Selhausen</b>	0-15 cm	15.4	67.5	17.1	1.03	0.069	0.504	0.0056	1.68	0.01
	15-33 cm	15.6	67.7	16.6	0.96	0.109	0.504	0.0059	1.92	0.05
	33-57 cm	16.2	63.1	23.1	0.34	0.000	0.463	0.0061	1.28	0.35
	57-120 cm	12.3	64.0	23.7	0.24	0.044	0.441	0.0013	1.69	0.05
<b>Merzenhausen</b>	0-12 cm	6.4	78.2	15.4	1.0	0.001	0.462	0.0031	1.69	0.30
	12-40 cm	6.4	78.2	15.4	1.0	0.001	0.571	0.0039	1.63	0.41
	40-60 cm	1.0	77.1	21.9	0.4	0.057	0.418	0.0034	1.21	0.64
	60-110 cm	0.5	73.4	26.1	0.3	0.103	0.367	0.0017	1.88	0.13
<b>Rollesbroich</b>	0-5 cm	22.0	60.8	17.2	4.82	0.034	0.443	0.0082	2.83	2.16
	5-14 cm	22.0	60.8	17.2	4.82	0.056	0.380	0.0077	2.84	2.04
	14-34 cm	23.1	59.1	17.8	2.49	0.039	0.379	0.0109	1.68	1.75
	34-60 cm	23.2	59.3	17.5	0.81	0.038	0.340	0.0160	1.33	0.84
	60-100 cm	23.2	59.3	17.5	0.0	0.037	0.375	0.0131	1.06	0.71

Tab. A.2:

Selection of most important fitted plant parameters for the calibration of the plant growth module of AgroC. (WW: winter wheat; WB: winter barley; GL: grassland; DVS: development stage; DM: dry matter).

	Selhausen	Merzenhausen				Rollesbroich
	WW 2009	WW 2012	WW 2013	WB 2014	GL 2013	
prescribed threshold pressure	-10,	-100,	-100,	-100,	-5,	
heads $h_0$ , $h_1$ , $h_2$ , and $h_3$ for scaling	-100,	-400,	-400,	-400,	-70,	
the root water uptake (cm)	-300,	-1000,	-1000,	-1000,	-150,	
	-800	-10000	-10000	-10000	-800	
specific leaf area of new leaves (ha leaf kg <sup>-1</sup> DM)	0.0024	0.0024	0.0023	0.0033	0.003	
potential CO <sub>2</sub> assimilation rate of a unit leaf area for light saturation (kg CO <sub>2</sub> ha <sup>-1</sup> leaf h <sup>-1</sup> )	47.0	60.0	53.0	48.0	75.0	
initial light use efficiency ((kg CO <sub>2</sub> ha <sup>-1</sup> leaf h <sup>-1</sup> )(J m <sup>-2</sup> s <sup>-1</sup> ) <sup>-1</sup> )	0.5	0.5	0.5	0.45	0.36	
DVS against reduction factor of the maximal light assimilation rate	0.0 1.0	0.0 1.0	0.0 1.0	0.0 1.0	0.0 1.0	
	1.0 1.0	1.0 1.0	1.0 1.0	1.0 1.0	0.4 1.0	
	2.0 0.4	2.0 0.5	2.0 0.4	2.0 0.3	1.0 0.9	
					1.2 0.9	
					1.5 0.9	
					1.8 0.9	
daily average daytime temperature against reduction factor of the maximal light assimilation rate	0.0 0.05	0.0 0.01	0.0 0.05	0.0 0.6	0.0 0.4	
	4.0 0.3	6.0 0.3	6.0 0.1	5.0 0.7	5.0 0.6	
	10.0 0.6	10.0 0.7	10.0 0.5	15.0 0.9	10.0 1.0	
	15.0 0.8	17.0 1.0	20.0 1.0	18.0 1.0	15.0 1.0	
	20.0 1.0	25.0 0.5	25.0 0.7	25.0 0.6	20.0 0.8	
	30.0 0.0	35.0 0.4	35.0 0.6	40.0 0.3	35.0 0.2	
DVS against fraction of dry matter allocated to the shoot	0.0 0.33	0.0 0.24	0.0 0.24	0.0 0.34	0.0 0.62	
	0.1 0.33	0.1 0.24	0.1 0.24	0.51 0.44	0.2 0.52	
	0.2 0.42	0.2 0.33	0.2 0.33	0.72 0.84	0.4 0.49	
	0.4 0.67	0.4 0.58	0.4 0.58	1.7 0.99	0.7 0.57	
	0.5 0.78	0.5 0.64	0.5 0.64	2.0 1.00	1.0 0.64	
	0.7 0.85	0.7 0.72	0.7 0.72		1.3 0.47	
	0.9 0.92	0.9 0.80	0.9 0.80		2.0 0.55	
	1.2 1.0	1.5 0.91	1.5 0.91			
	2.0 1.0	2.0 1.0	2.0 1.0			

## 1 **Appendix B: Results and Discussions Supporting Figures**

2

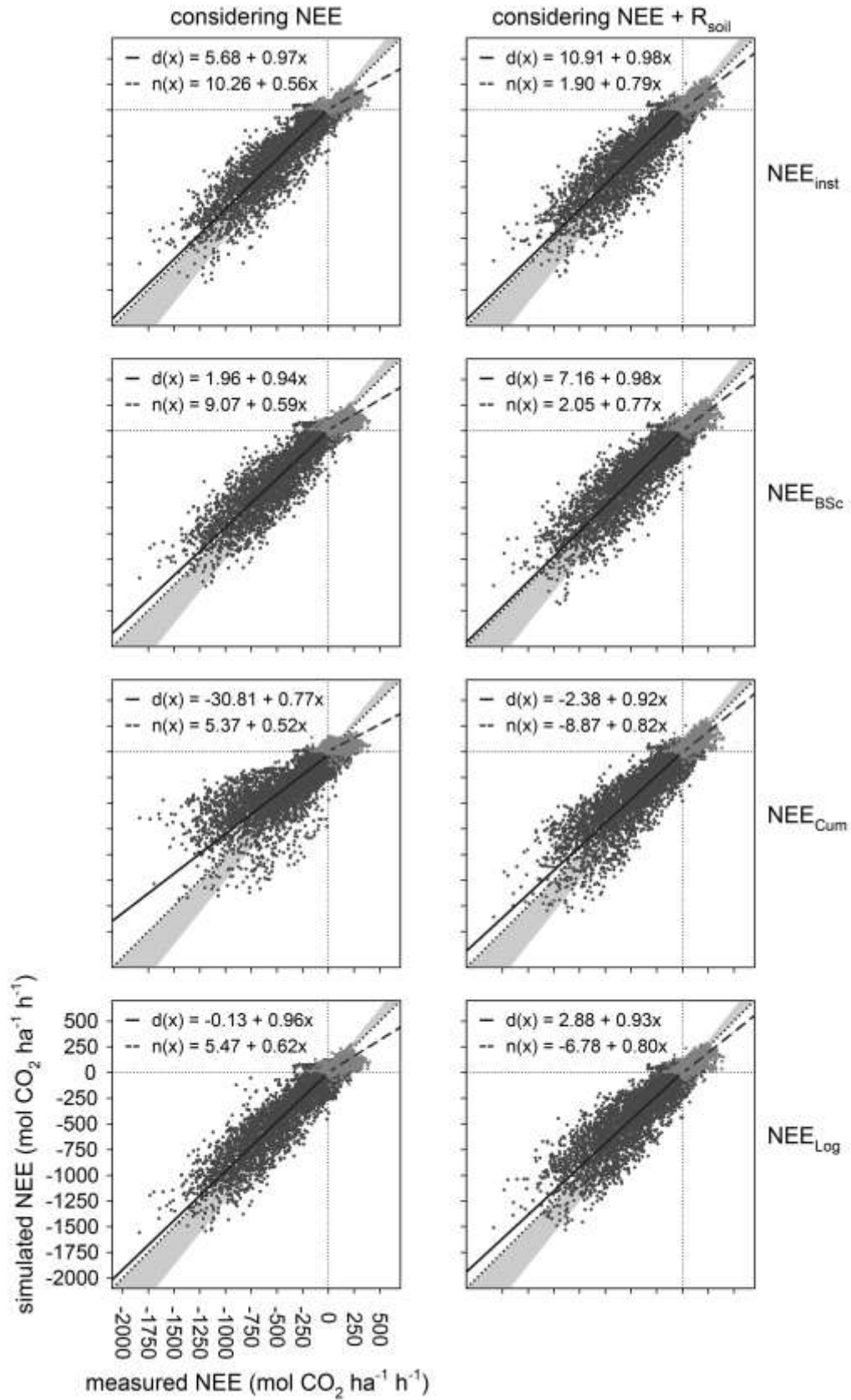
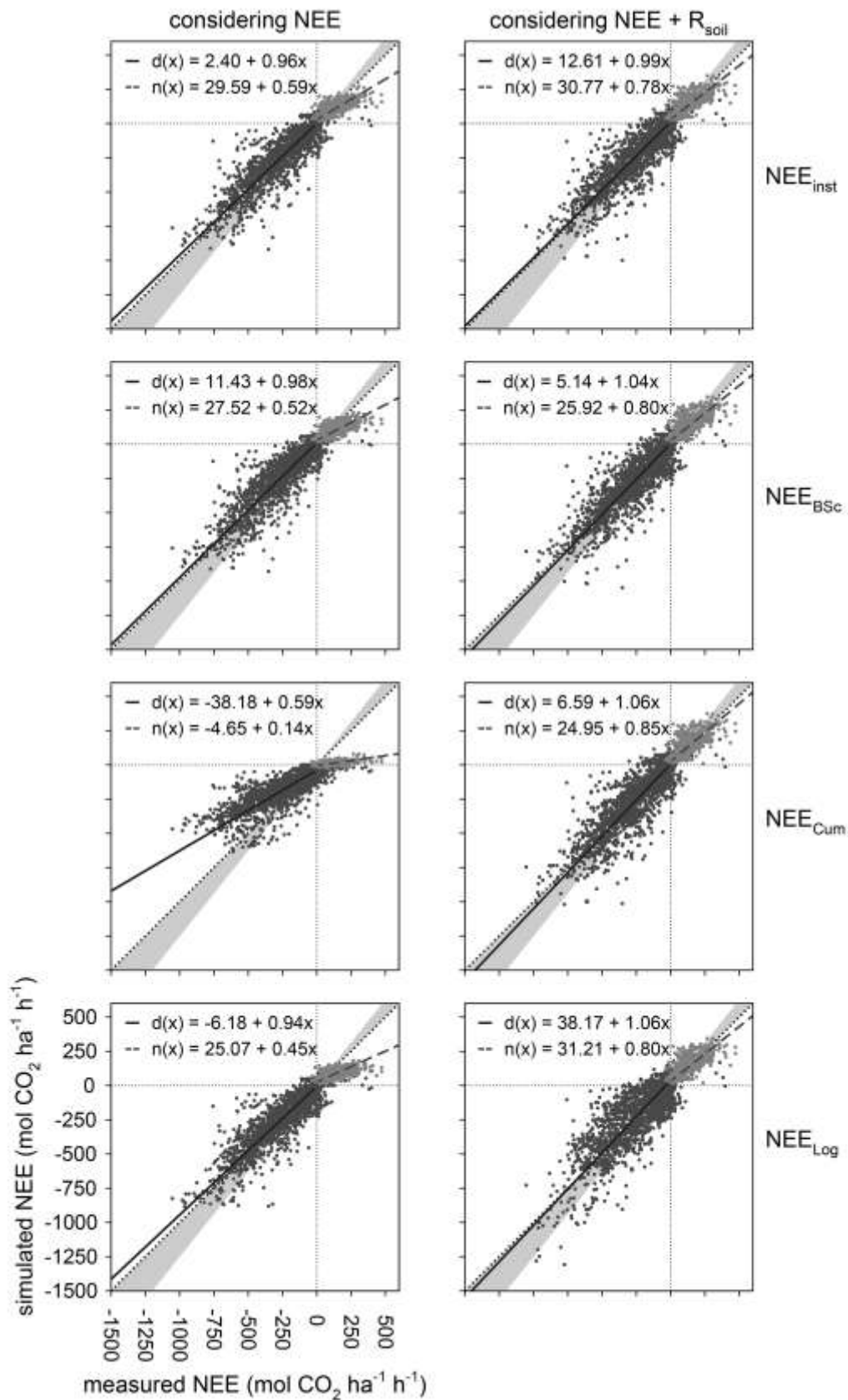
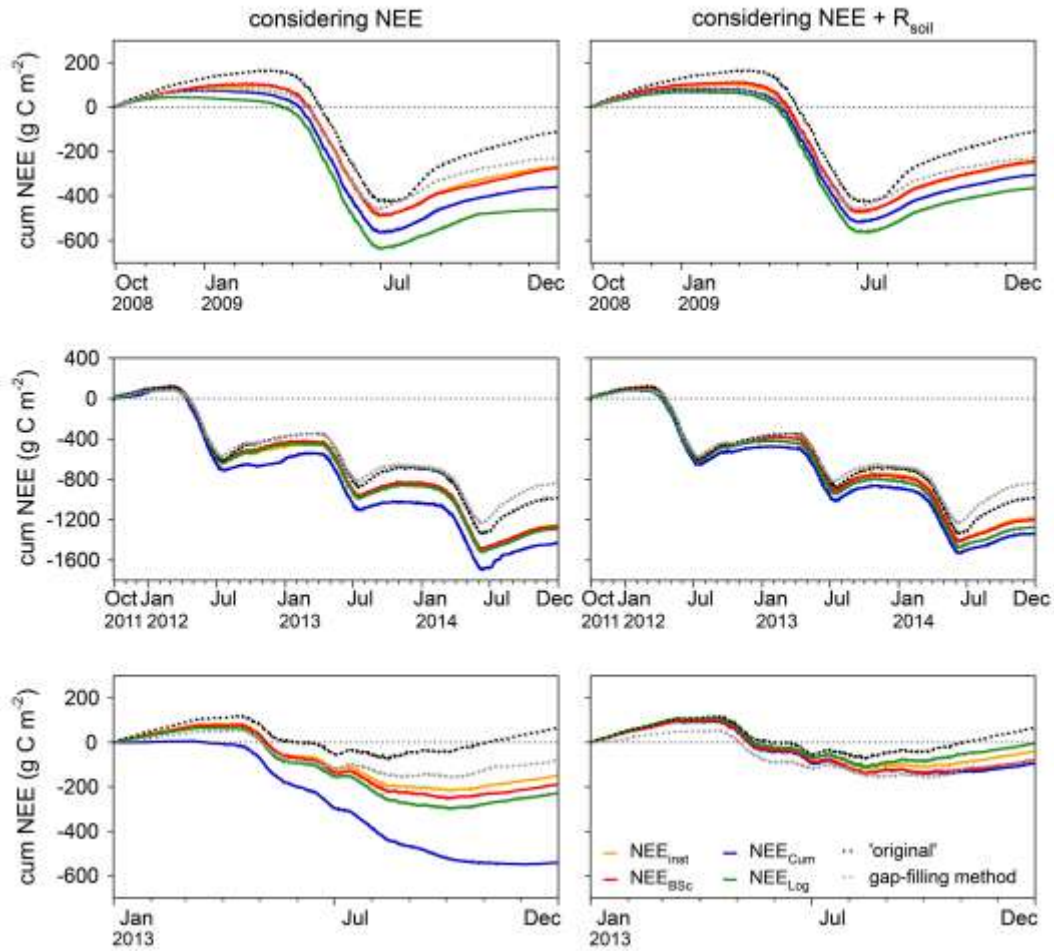


Fig. B.1:

Correlations between observed and simulated net ecosystem exchange (NEE) for all optimization strategies at test site Merzenhausen. Reduced major axis regression was derived for each strategy distinguished between day- (d) and nighttime (n) CO<sub>2</sub> fluxes, whereat nighttime was designated to a measured global radiation lower than 20 W m<sup>-2</sup>. For description of optimization strategies see text.



1 *Fig. B.2:*  
2 Correlations between observed and simulated net ecosystem exchange (NEE) for all  
3 optimization strategies at test site Rollesbroich. Reduced major axis regression was derived  
4 for each strategy distinguished between day- (d) and nighttime (n) CO<sub>2</sub> fluxes, whereat  
5 nighttime was designated to a measured global radiation lower than 20 W m<sup>-2</sup>. For description  
6 of optimization strategies see text.



1 *Fig. B.3:*  
2 Cumulated net ecosystem exchange (cum NEE) over simulation time period, calculated in  
3 “gap-filling mode”, for each optimization strategy, for the simulation without calibration to  
4 NEE (‘original’), and for the gap-filling method after Reichstein et al. (2005) (gap-filling  
5 method) in Selhausen (top), Merzenhausen (middle), and Rollesbroich (bottom). For  
6 description of optimization strategies see text.